

# Mass Spectrometric Methods for Predictive Characterization of Lyophilized Products

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# Amorphous solids and pharmaceutical biotechnology

- ▶ Recombinant protein drugs are often stored as amorphous powders.
  - Bulk API storage
  - Final product
- ▶ Of nine protein drug products approved by US FDA in 2011, five are solids.
- ▶ Produced by lyophilization or spray drying.
- ▶ Degradation can occur during lyophilization and storage.
- ▶ Few analytical methods for protein conformation in solids, low resolution, semi-quantitative.


# Analytical methods for protein structure

- In solution

- NMR (2-D)
- Circular dichroism (CD)
- Infrared (FTIR)
- NIR
- Raman
- Fluorescence
- U.V. absorption
- H/D exchange
- X-ray crystallography

- In solids

- FTIR
- NIR
- Raman
- Fluorescence
- ssNMR
- H/D exchange



*Low resolution  
Semi-quantitative*

# Central hypothesis

*Chemical definition* of amorphous solids is possible and can be used to rationally design solid formulations of protein drugs.

## Current paradigm:

- chemical environment in amorphous solids is unknowable,
- protein conformation in solids can only be measured with low resolution,
- trial-and-error informed by limited physical characterization is the only possible approach to formulation development.

# Hydrogen/deuterium exchange (HDX) for proteins in solids

» Conformation and excipient interactions with peptide-level resolution

Y. Li et al., *Biotech. Bioeng.*, 97/6: 1650–1653, 2007

Y. Li et al., *Anal. Biochem.*, 366:18–28, 2007

Y. Li et al. *Pharm. Res.*, 25/2: 259–267, 2008

S. Sinha et al., *Biophys. J.*, 95/12: 5951–5961, 2008

A. Sophocleous et al., *Mol. Pharm.*, 9/4: 718–726, 2012

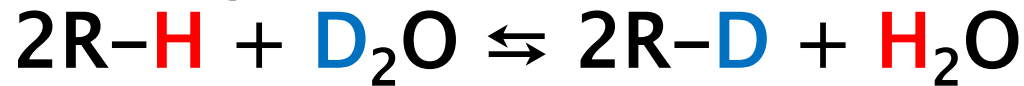
A. Sophocleous et al., *Mol. Pharm.*, 9/4: 727–733, 2012

B.S. Moorthy et al., *Mol. Pharm.*, 11/6: 1869–1879, 2014

B.S. Moorthy et al., *JoVE*, (98), e52503, 2015

# What is HDX?

- ▶ A chemical reaction, in which labile hydrogen atoms “exchange” with deuterium.

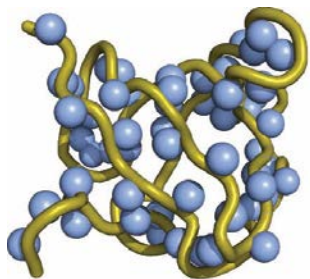


- ▶ Reaction is quenched at low pH and low temperature.
- ▶ The increase in mass (+1 amu) can be detected using mass spectrometry (LC-MS, LC-MS/MS).
- ▶ The rate and extent of HDX provides information on protein structure and dynamics in aqueous solution.
- ▶ Due to back exchange, only deuteration of backbone amide groups is detected.



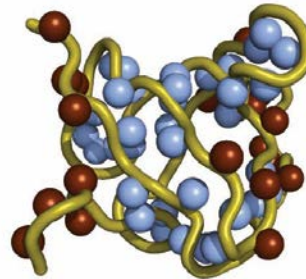
# Basic principles of HDX

## H/D Exchange



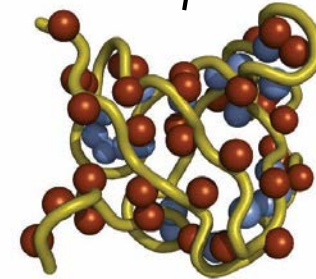
● H ● D

$D_2O$   
Time



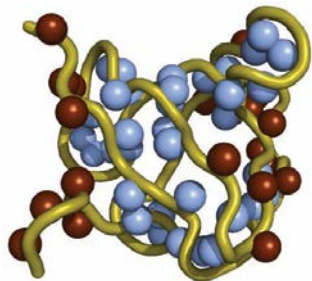
Dynamic regions  
exchange rapidly

$D_2O$   
Time

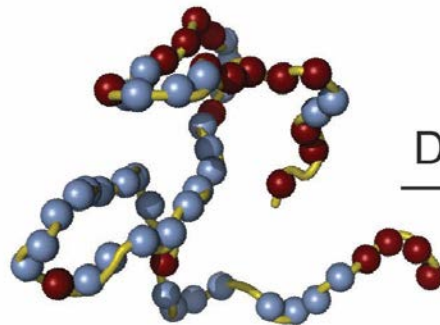


Structured regions  
exchange slowly

## Quench and Digest

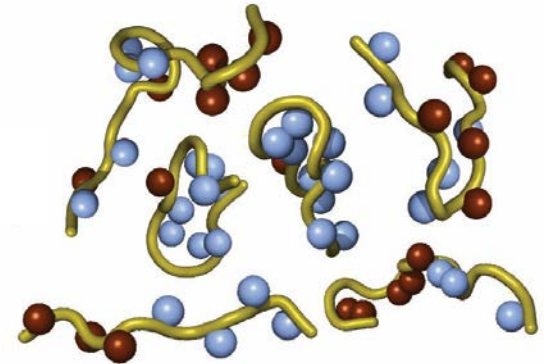


Quench  
pH 2.5  
0 °C



Quenching locks in deuterium  
and unfolds the protein

Digest

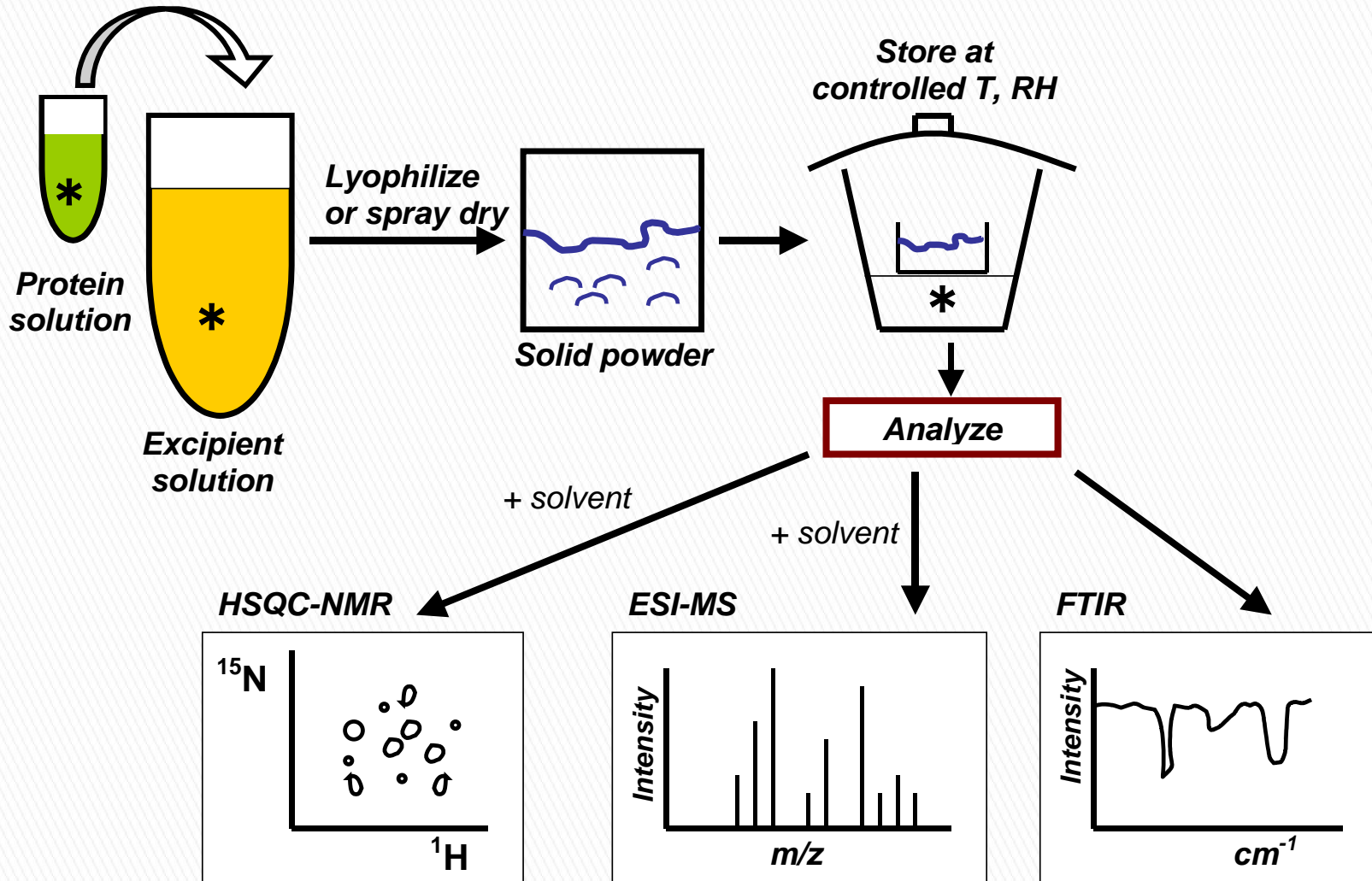


Digestion localizes the  
information

MS analysis

# HDX for lyophilized solids (ssHDX)

Expose powder to D<sub>2</sub>O vapor





# Potential benefits of ssHDX

...and some unknowns

## *POTENTIAL BENEFITS*

- ▶ Information on protein structure, with **peptide-level** resolution.
- ▶ **Screen** formulations and/or processes for effects on protein structure.
- ▶ Obtain fundamental information on protein **structure and dynamics** in amorphous solids.
- ▶ Obtain fundamental information on **amorphous solids** (spatial, dynamic heterogeneity) with protein as a reporter.

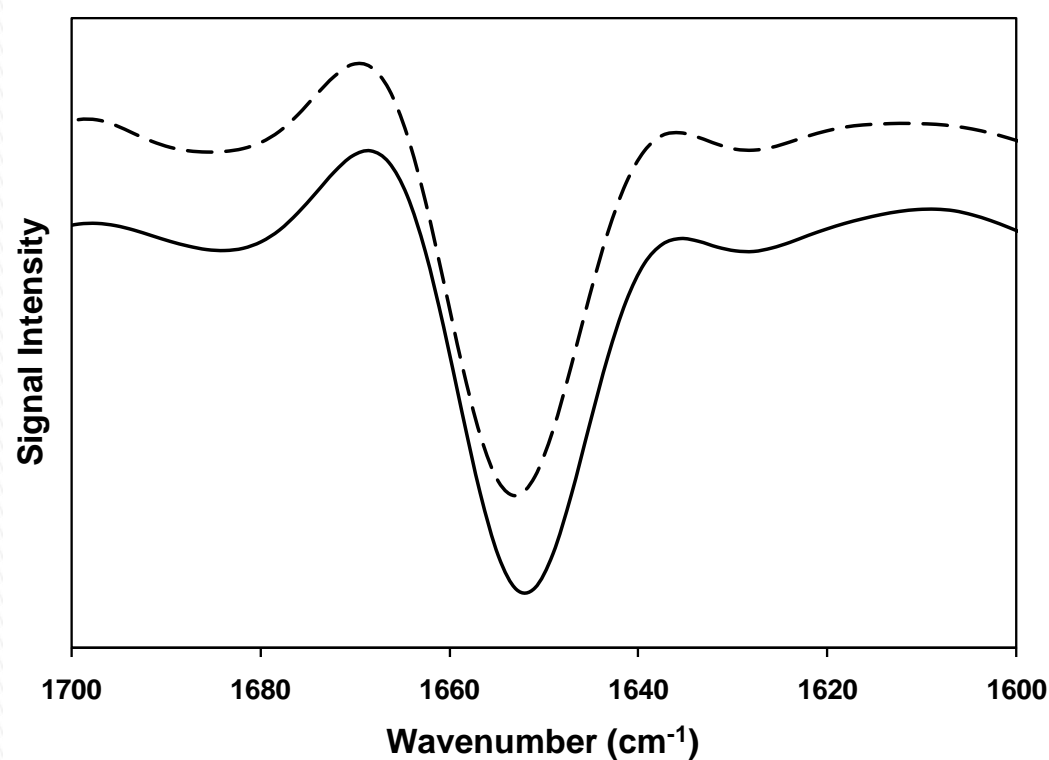
## *UNKNOWNNS*

- ▶ Will it **work**?
- ▶ Will D<sub>2</sub>O(g) **sorption** kinetics affect the results?
- ▶ Are HDX results predictive of storage stability?
- ▶ How should the results be interpreted?

*Myoglobin (holo)*



# No detectable secondary structural differences observed post-lyophilization by FTIR

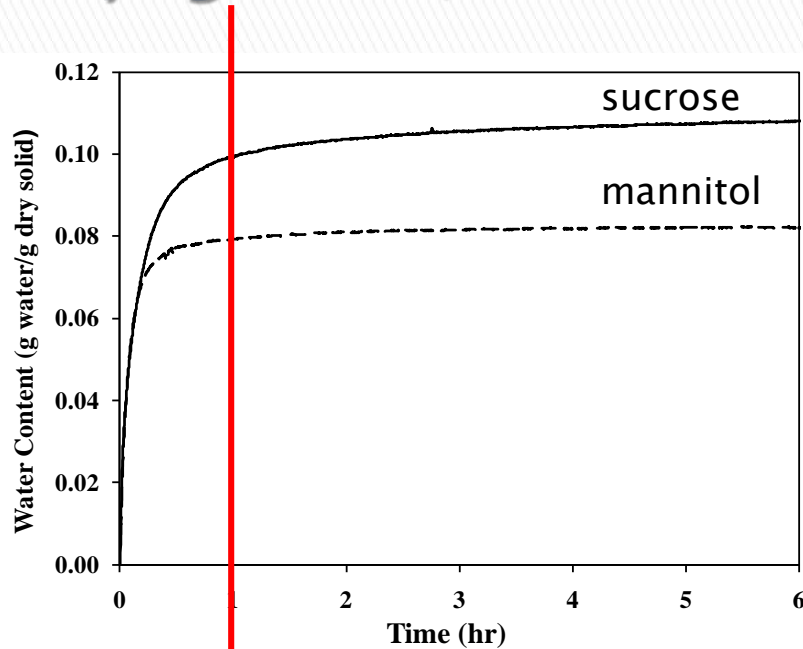


native-like  $\alpha$ -helix  
peak at  $1655 \text{ cm}^{-1}$

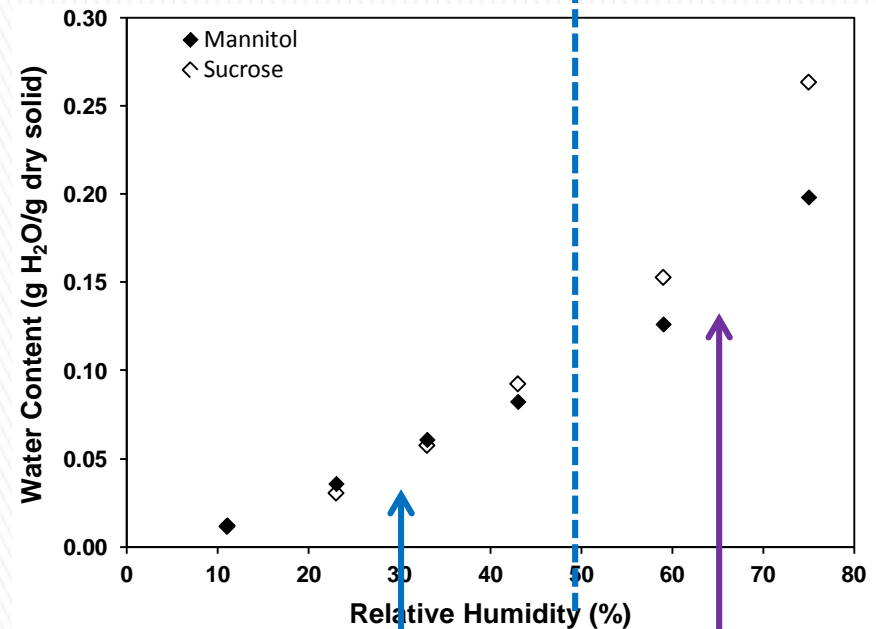
**Second-derivative FTIR spectra of Mb in lyophilized solids containing mannitol (solid line) or sucrose (dashed line).**

# Water sorption and water content

## *Myoglobin / mannitol or sucrose 1:1, 5 °C*



Water vapor sorption is essentially complete in ~ 1 h.

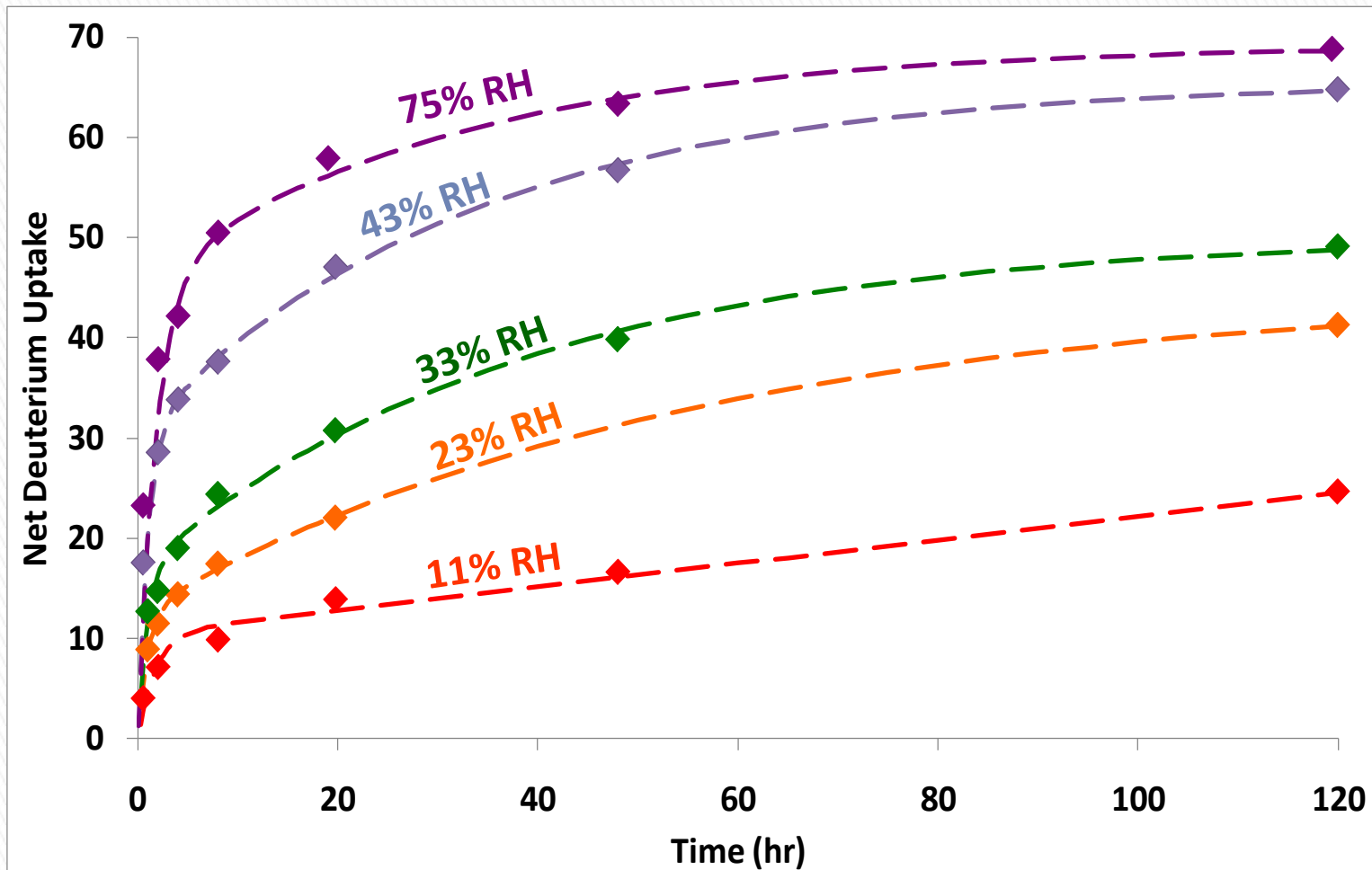


At low RH, water content of mannitol and sucrose formulations are equal.

At high RH, water content in sucrose formulation is greater than in mannitol.

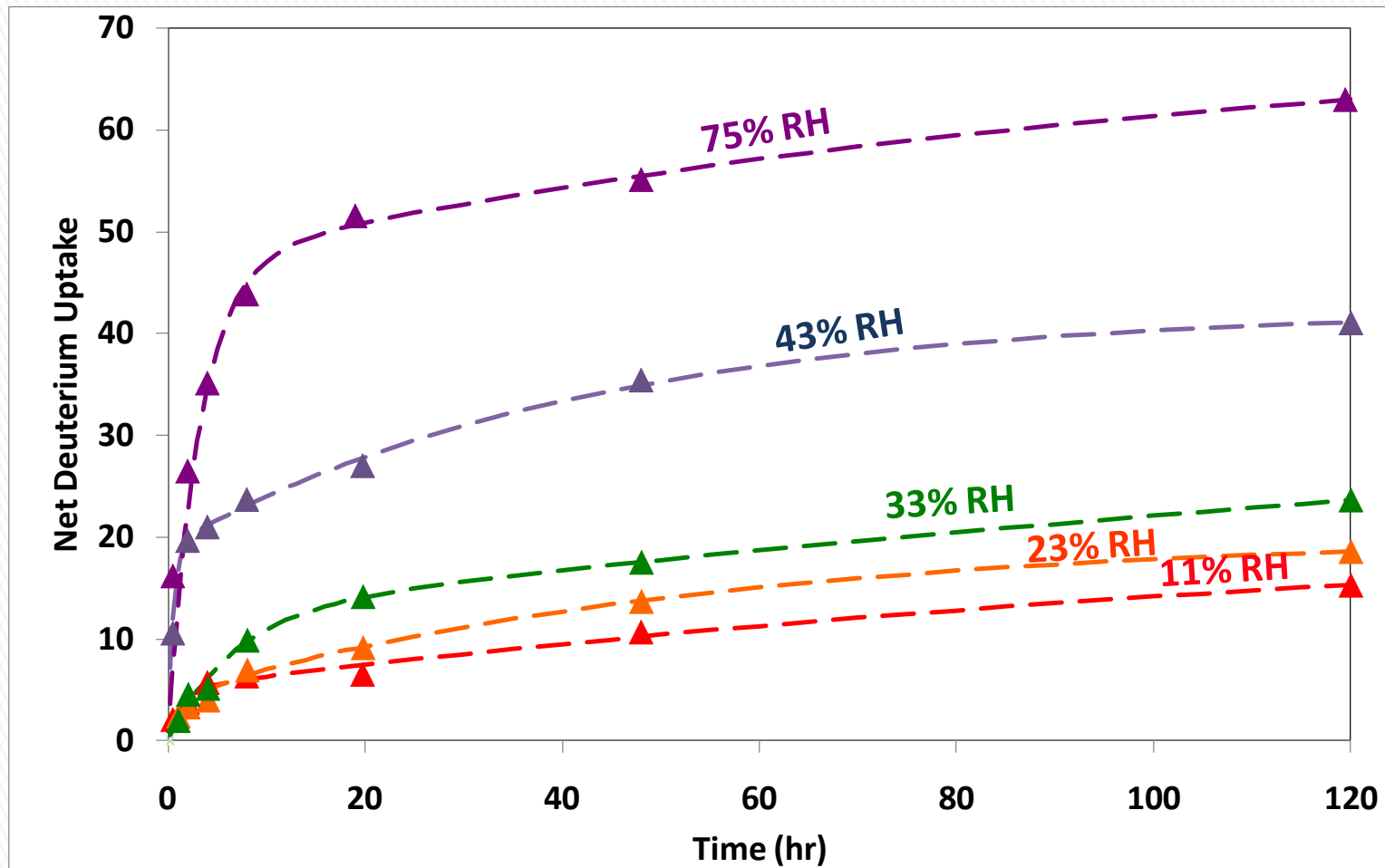
# Effect of RH on deuterium uptake

Intact protein: *Myoglobin* / *mannitol* 1:1, 5 °C



# Effect of RH on deuterium uptake

Intact protein: *Myoglobin* / *sucrose* 1:1, 5 °C



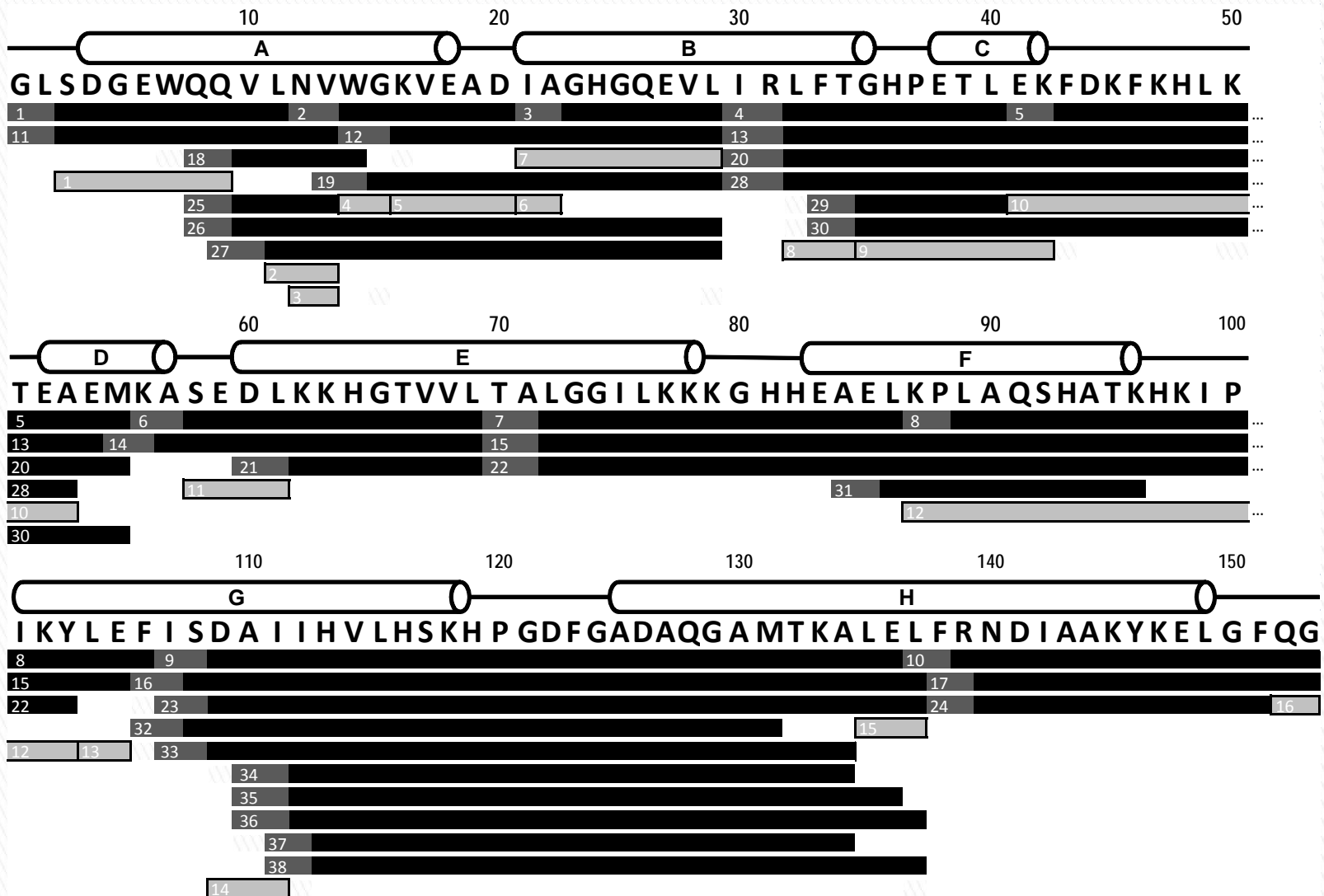
HDX in sucrose is less than mannitol  
*despite* similar or greater water content



protein structure/dynamics or spatial  
heterogeneity in water content

# Myoglobin peptide map

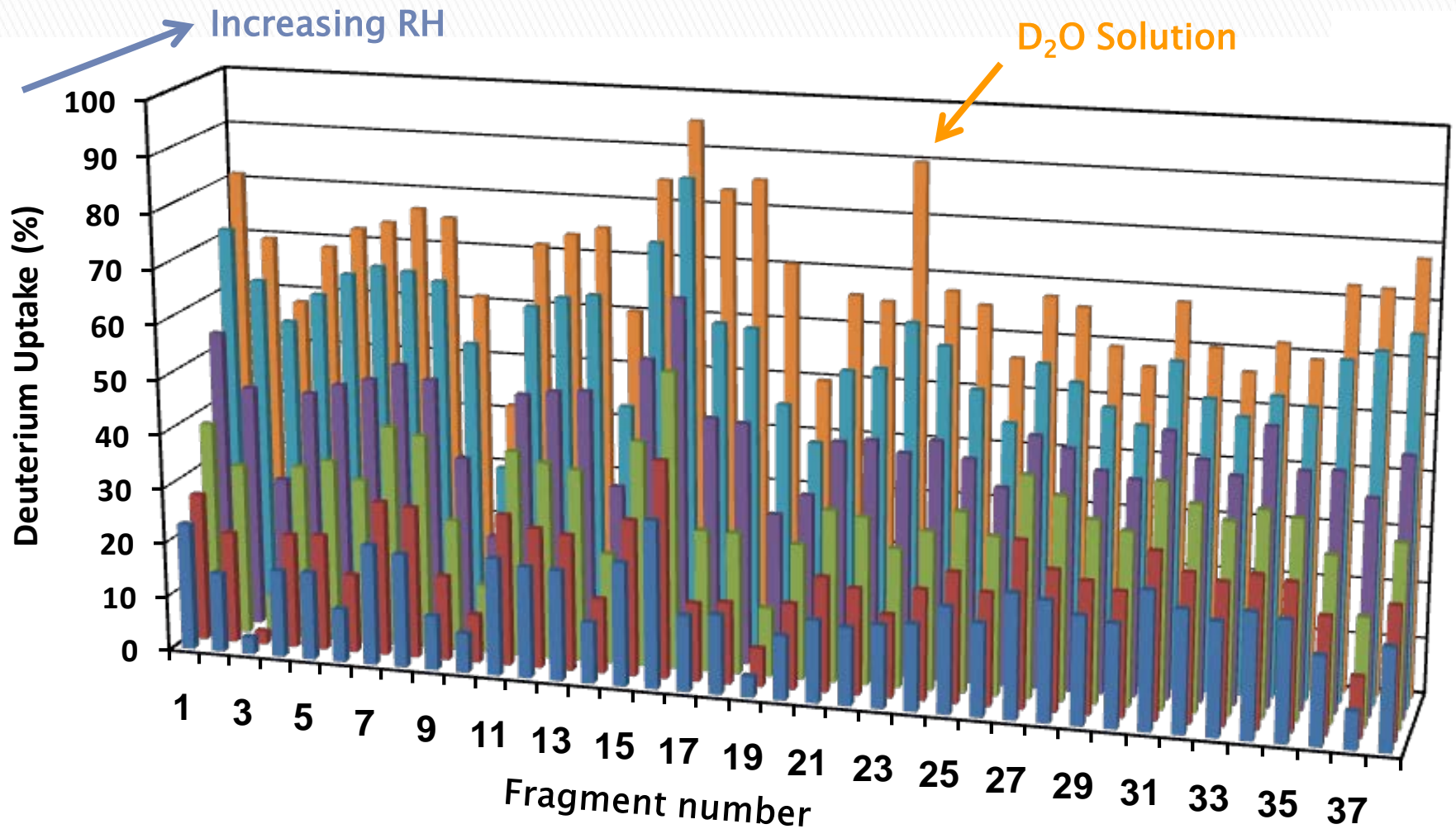
Pepsin digestion





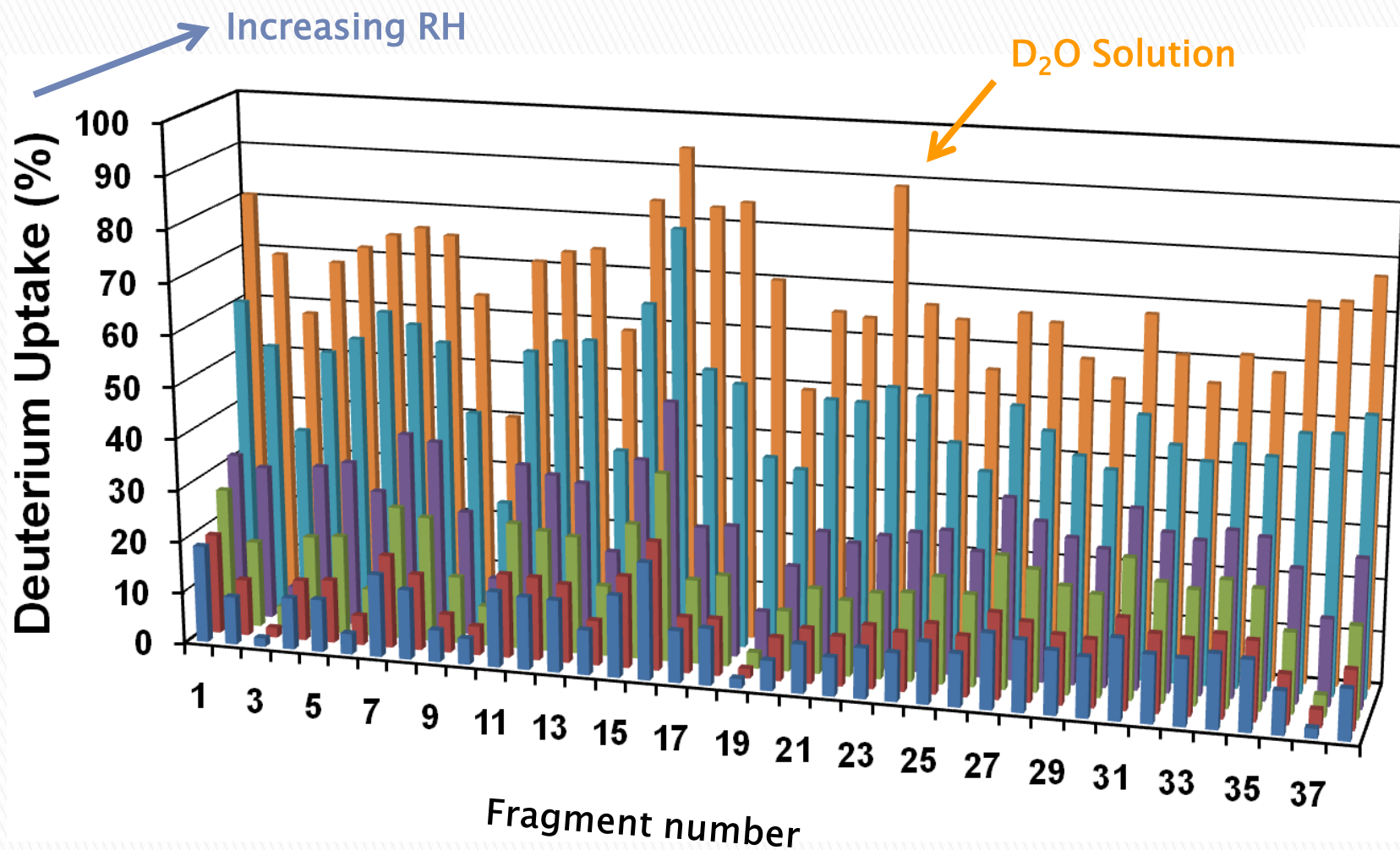
# Peptic fragments

*Myoglobin / mannitol 1:1, 5 °C*



# Peptic fragments

*Myoglobin / sucrose 1:1, 5 °C*



# Myoglobin in D<sub>2</sub>O solution

5 °C

Deuterium uptake

91-100%

81-90%

71-80%

61-70%

51-60%

41-60%

31-40%

21-30%

11-20%

0-10%



# Lyophilized Myoglobin: Mannitol

(1:1 w/w), 43% RH, 5 °C

Deuterium uptake

91-100%

81-90%

71-80%

61-70%

51-60%

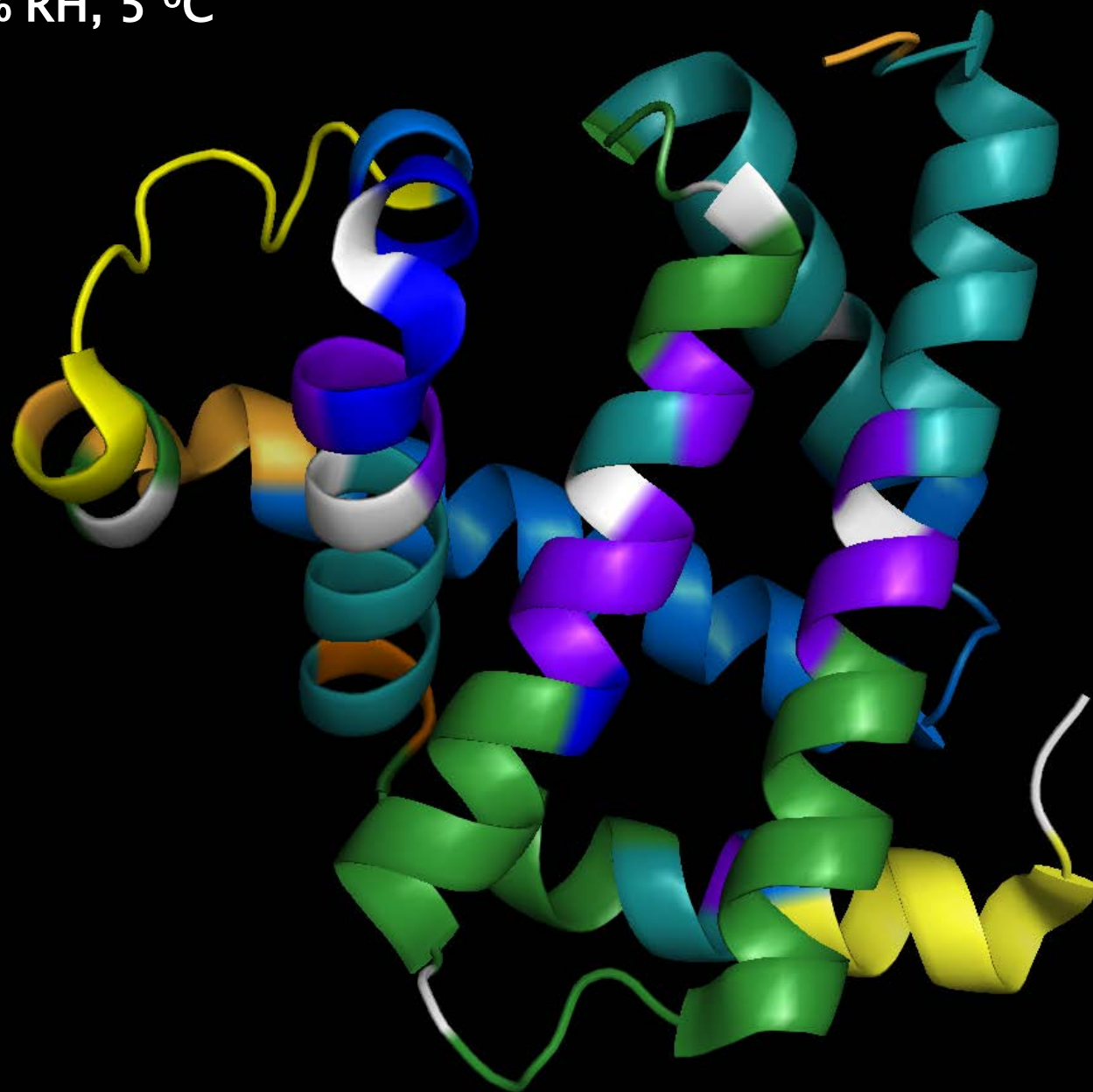
41-60%

31-40%

21-30%

11-20%

0-10%





# Lyophilized Myoglobin: Sucrose

(1:1 w/w), 43% RH. 5 °C

Deuterium uptake

91-100%

81-90%

71-80%

61-70%

51-60%

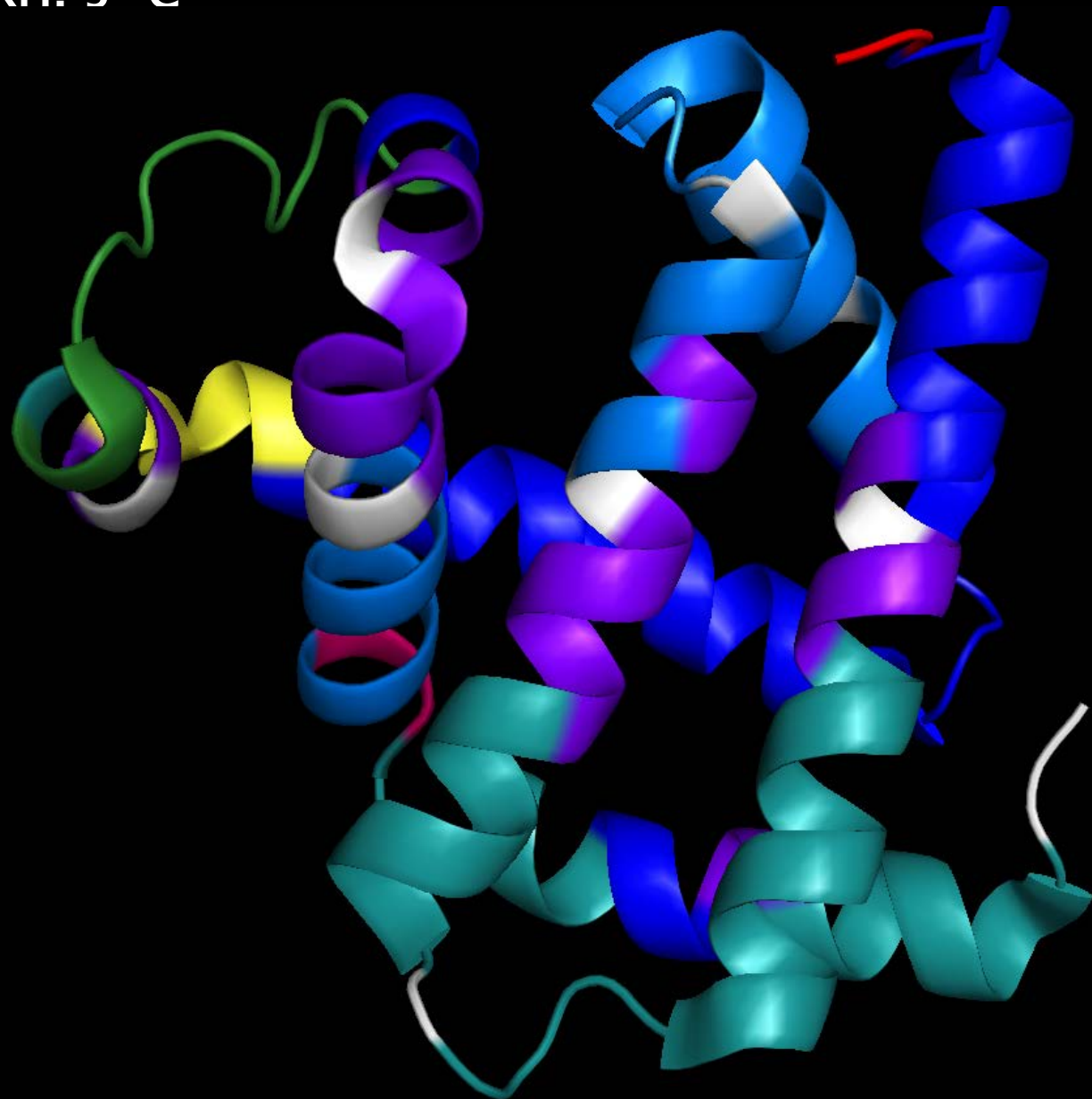
41-60%

31-40%

21-30%

11-20%

0-10%

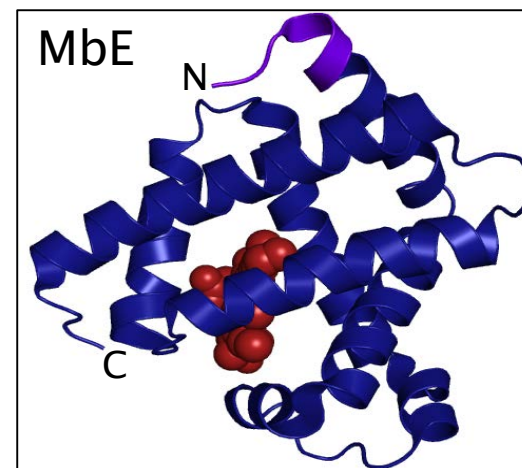
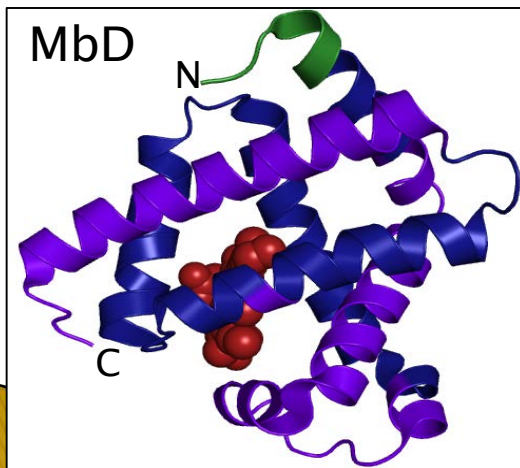
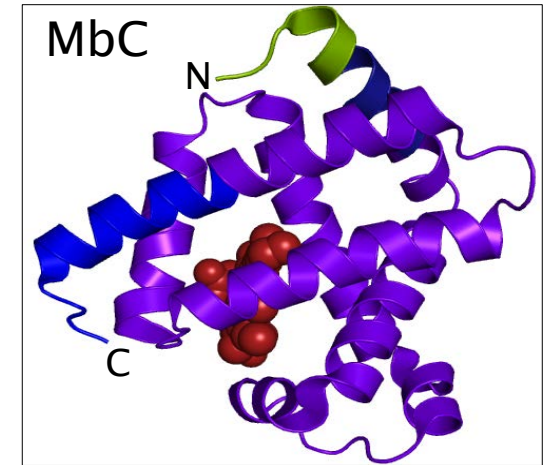
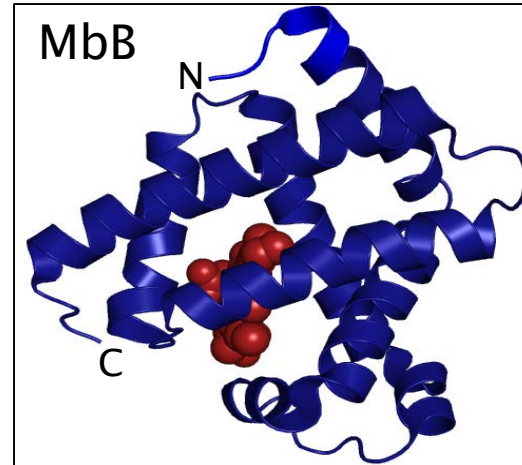
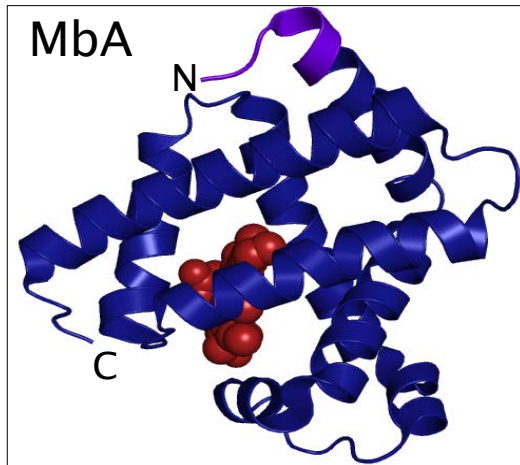


# ssHDX and Storage Stability

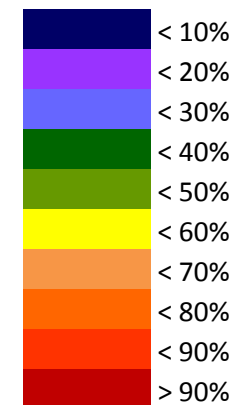
Five Mb formulations, one year storage at 25 and 40C

Ingredients	Formulations				
	MbA	MbB	MbC	MbD	MbE
Myoglobin (Mb)	1.7 mg/mL, 45% w/w	1.7 mg/mL, 45% w/w	1.7 mg/mL, 45% w/w	3.4 mg/mL, 90% w/w	0.4 mg/mL, 10% w/w
Sucrose	1.7 mg/mL, 45% w/w	-	-	-	3.0 mg/mL, 80% w/w
Mannitol	-	1.7 mg/mL, 45% w/w	-	-	-
NaCl	-	-	1.7 mg/mL, 45% w/w	-	-
Potassium Phosphate, (pH 7.0)	0.4 mg/mL, 10% w/w	0.4 mg/mL, 10% w/w	0.4 mg/mL, 10% w/w	0.4 mg/mL, 10% w/w	0.4 mg/mL, 10% w/w

# HDX Results Mapped on to the Mb Structure (T=1 h)

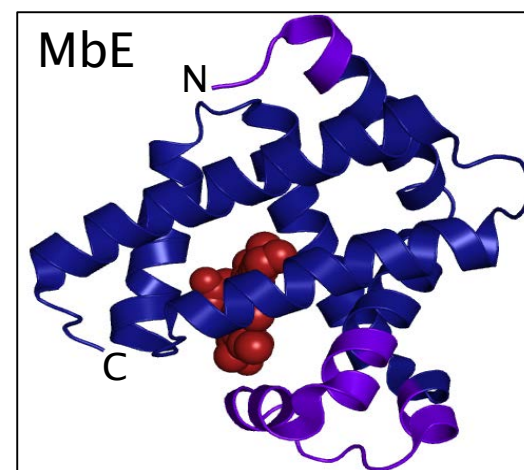
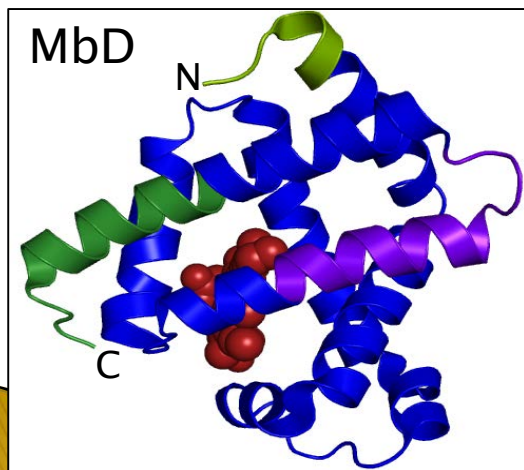
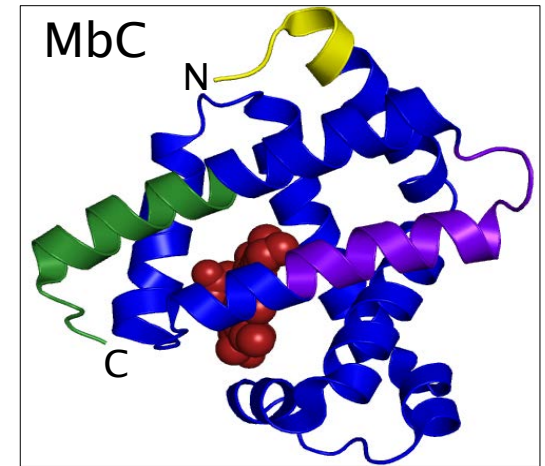
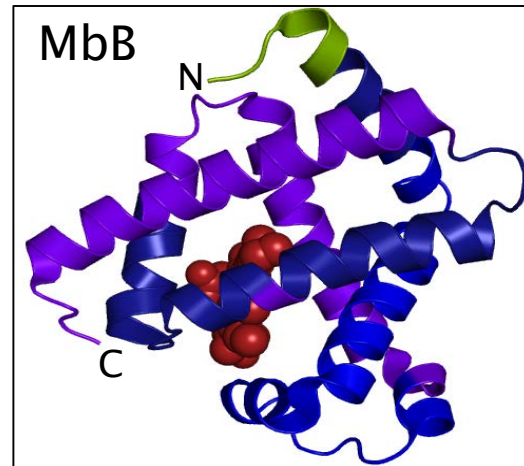
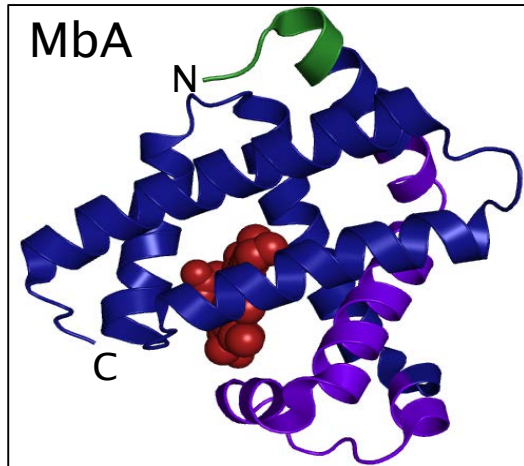


Deuterium Level

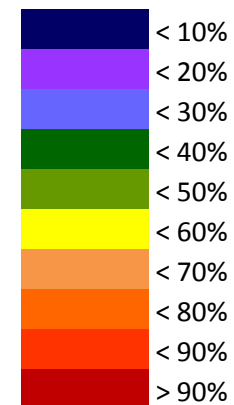




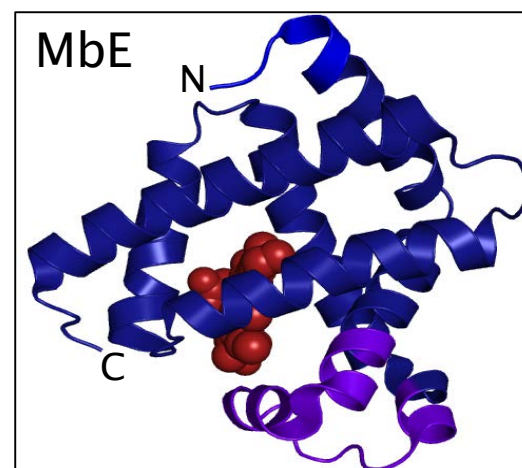
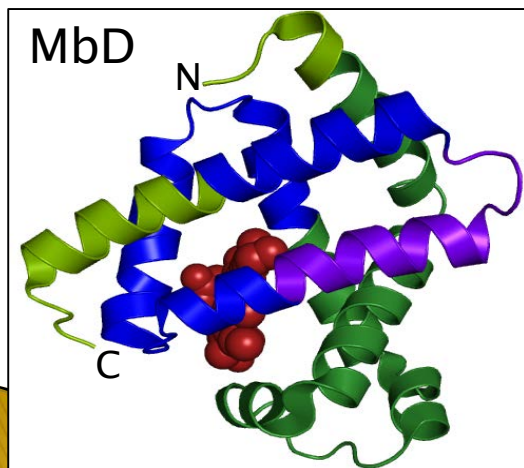
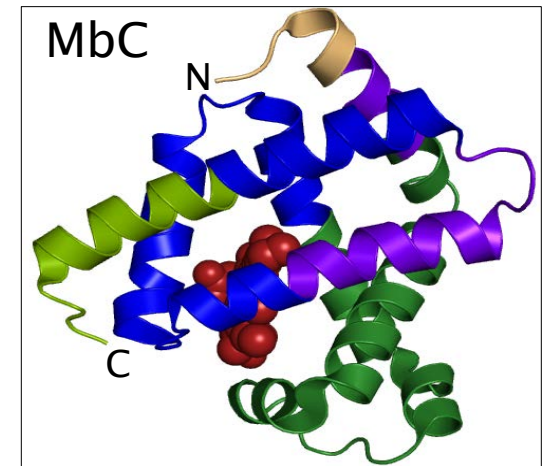
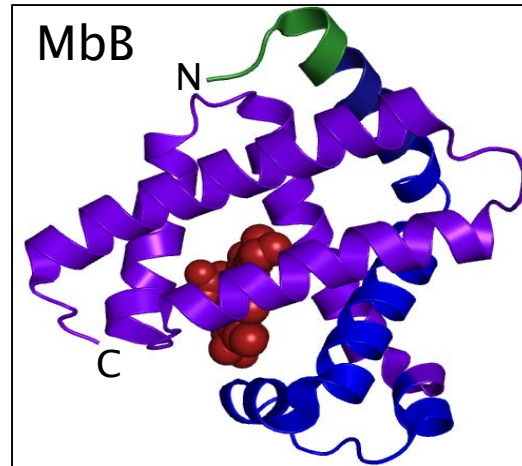
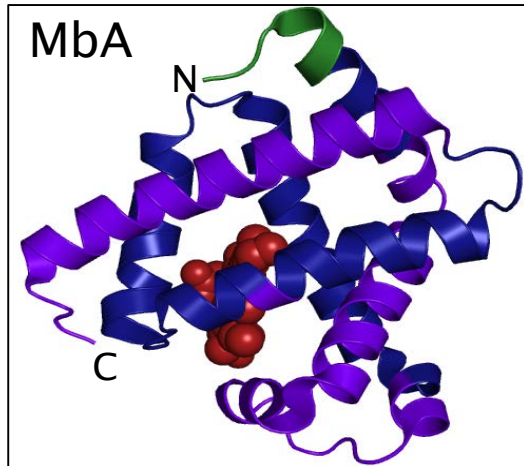
# HDX Results Mapped on to the Mb Structure (T=6h)



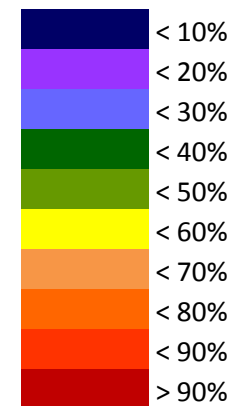
Deuterium Level



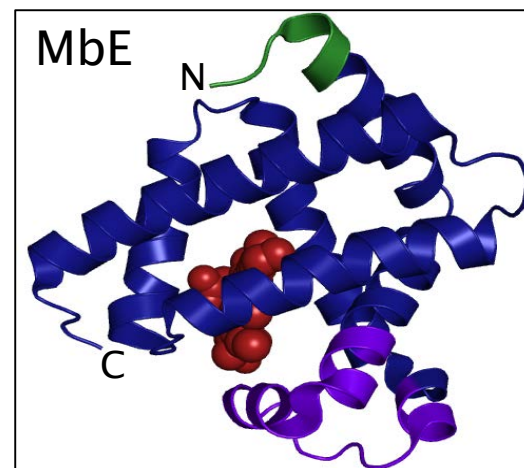
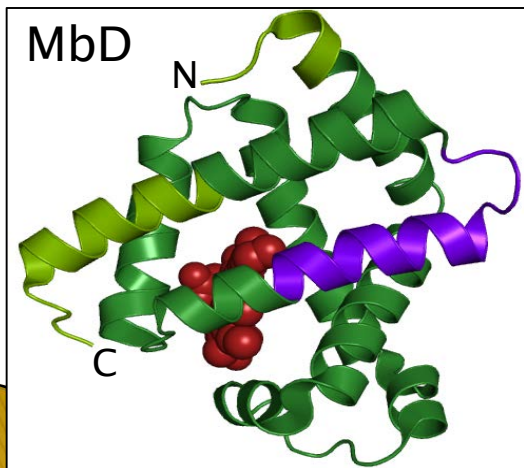
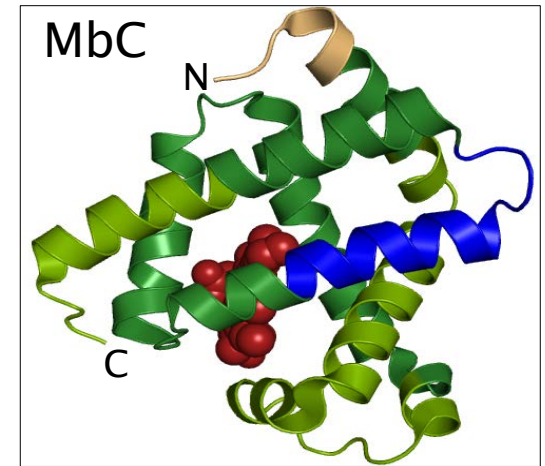
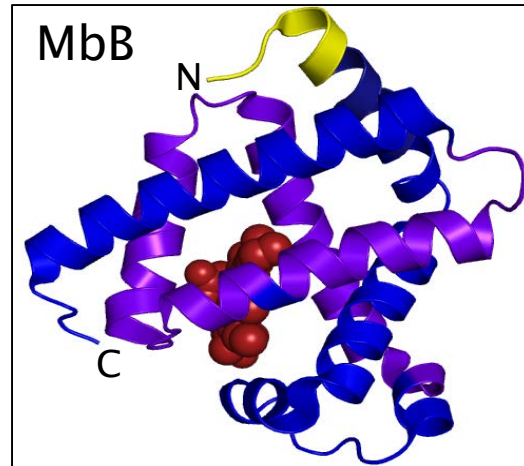
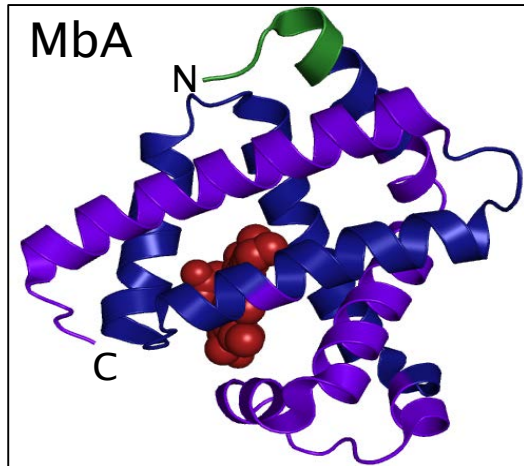
# HDX Results Mapped on to the Mb Structure (T=24h)



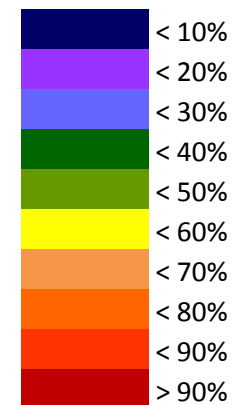
Deuterium Level



# HDX Results Mapped on to the Mb Structure (T=48h)

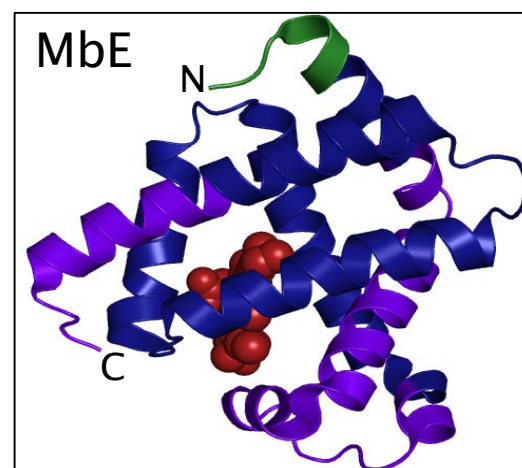
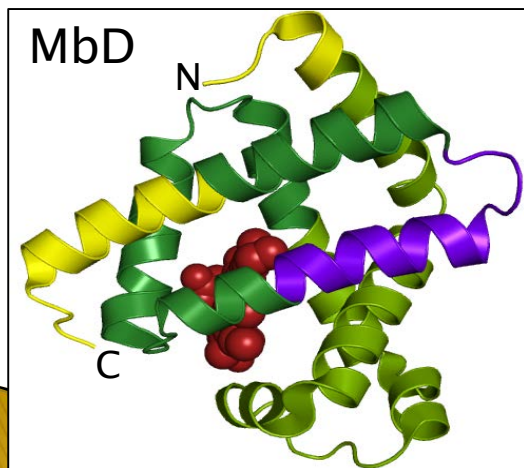
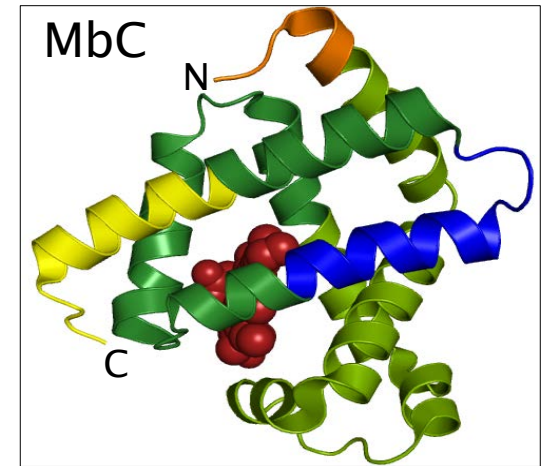
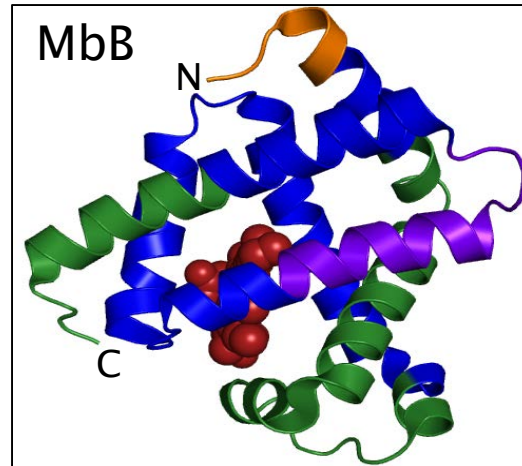
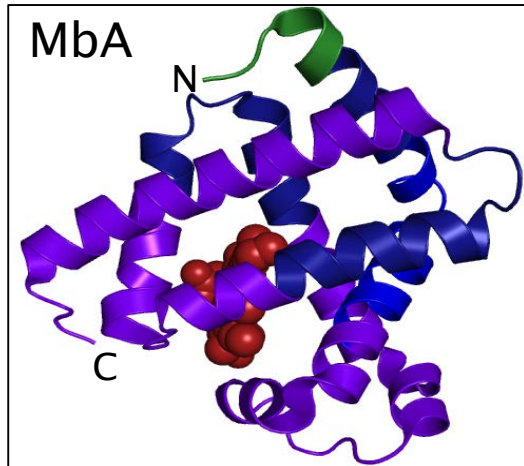


Deuterium Level

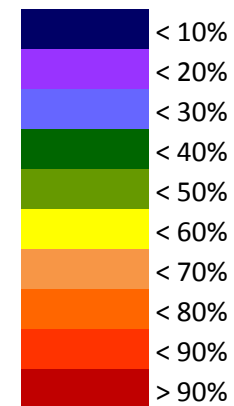




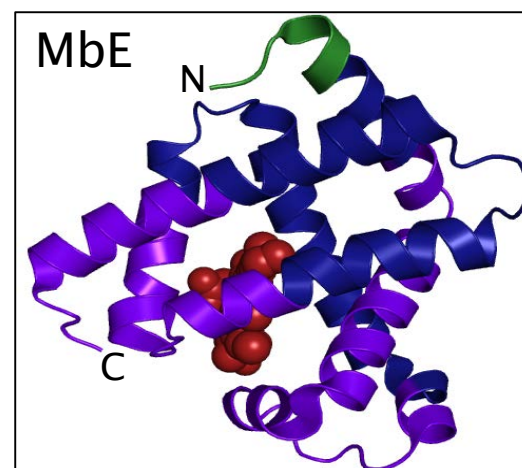
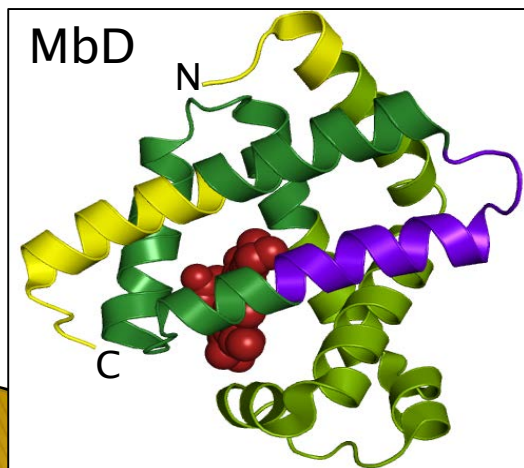
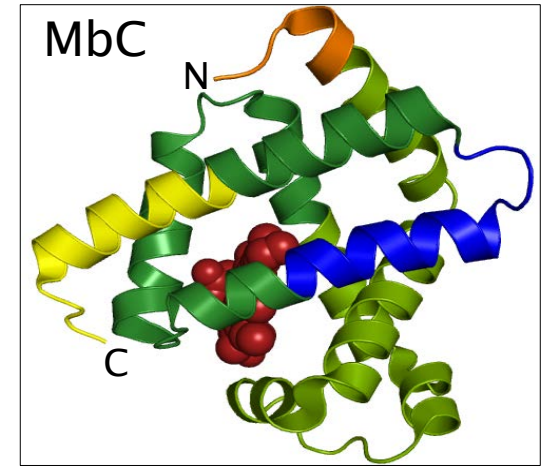
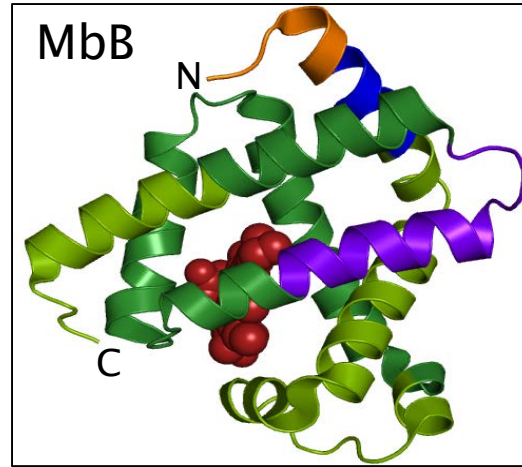
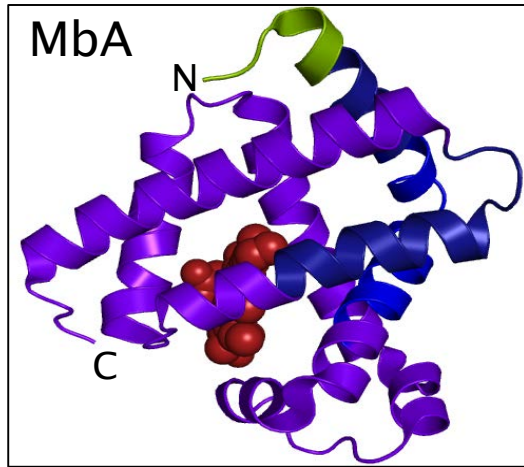
# HDX Results Mapped on to the Mb Structure (T=120h)



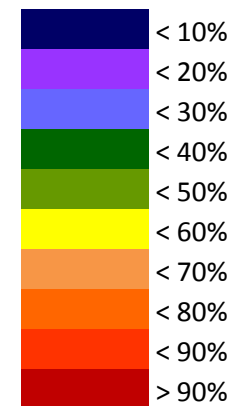
Deuterium Level



# HDX Results Mapped on to the Mb Structure (T=240h)

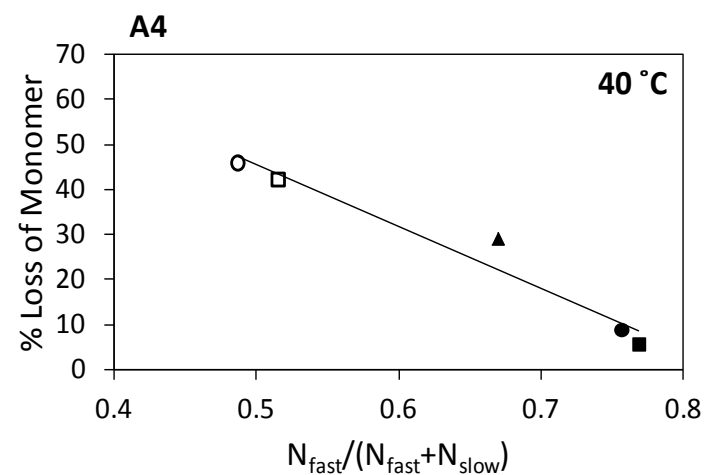
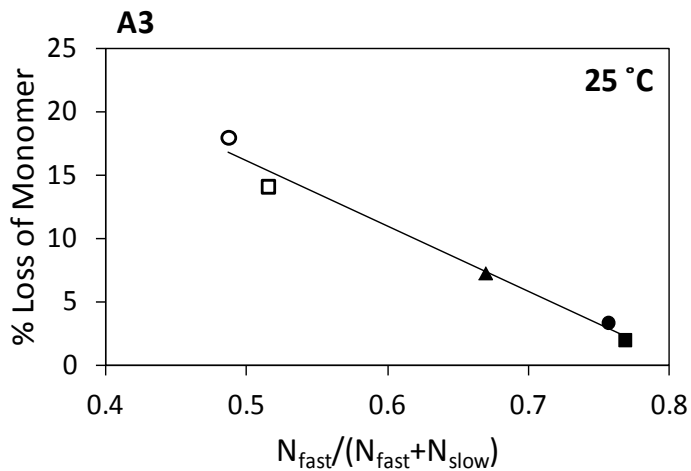
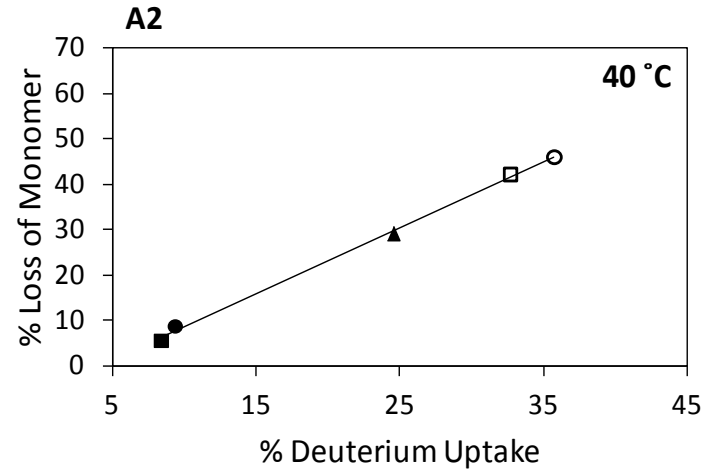
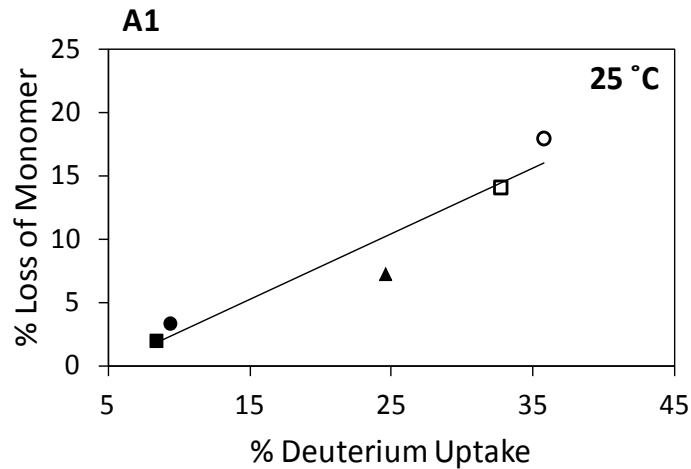


Deuterium Level



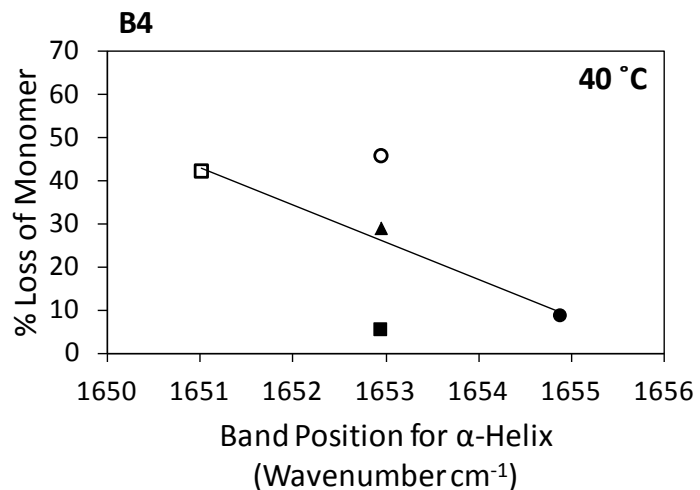
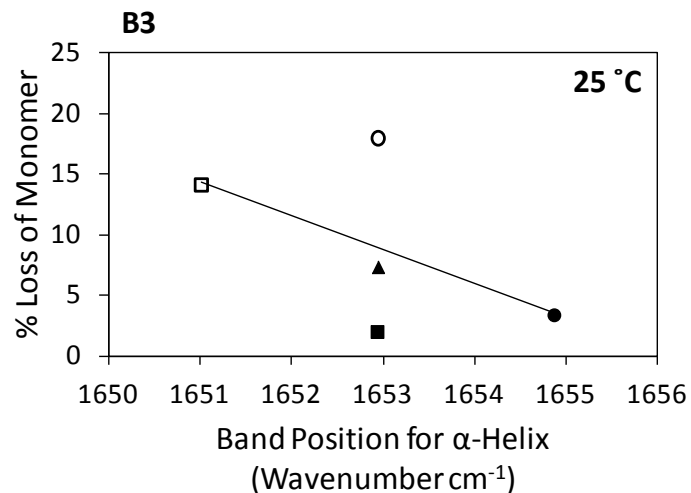
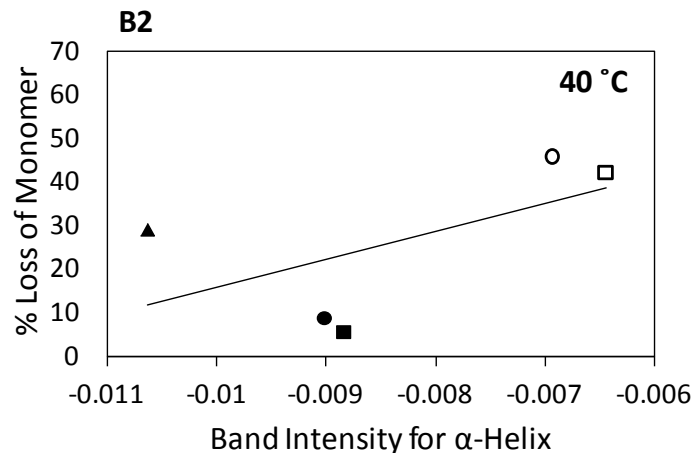
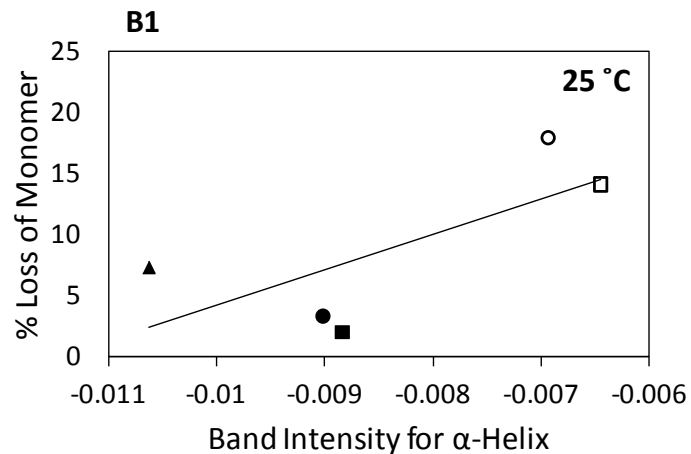
# ssHDX and storage stability

Storage for 360 days, 25 and 40 °C; stability by SEC; 48 h ssHDX



# FTIR and storage stability

Storage for 360 days, 25 and 40 °C; stability by SEC





# Summary

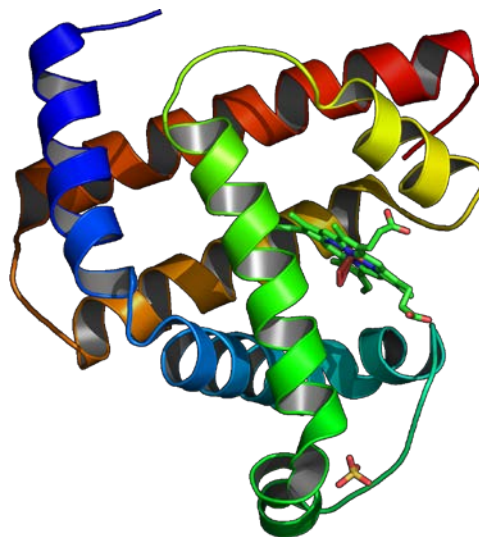
- ▶ ssHDX can be performed in powders using  $D_2O(g)$  as a deuterium source.
- ▶ ssHDX rates are much slower than vapor sorption.
- ▶ Rate and extent of ssHDX are affected by RH, excipient type and amount, temperature.
- ▶ For Mb in lyo powders, ssHDX parameters are highly correlated with extent of aggregation on 1-yr storage.

# Unknown

Relative contributions of protein conformation, protein dynamics and excipient interactions to ssHDX results.

## **Solution HDX**

- Solvent exposure
- Conformation
- Dynamics



## **Solid-state HDX**

- D<sub>2</sub>O exposure
- Conformation
- Dynamics; local?
- Matrix / excipient interactions?

# Solid-state photolytic labeling (ssPLL)

- » Mapping protein-protein interactions and the side-chain environment of proteins in amorphous solids

L. Iyer et al., *Mol. Pharm.*, 10: 4629–4639, 2013

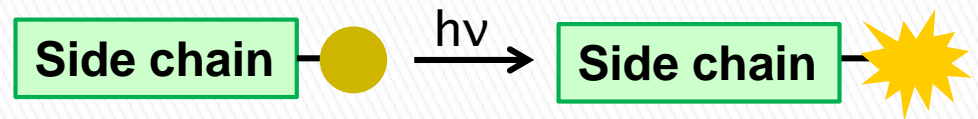
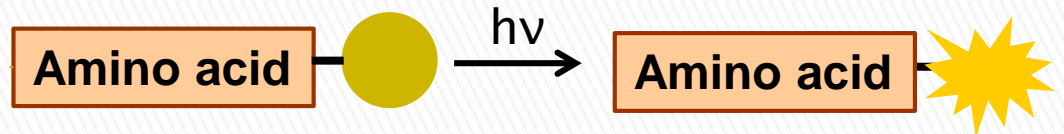
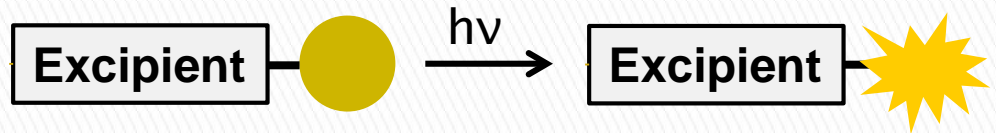
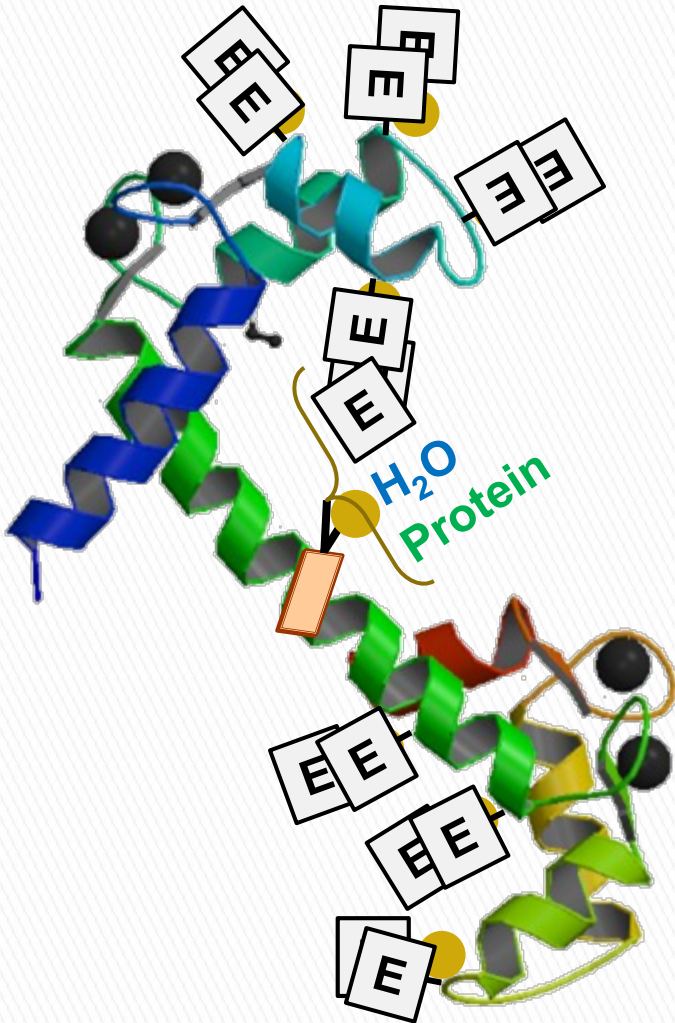
B.S. Moorthy et al., *JoVE*, in press

L. Iyer et al., *Mol. Pharm.*, in preparation

# Covalent labeling with MS analysis

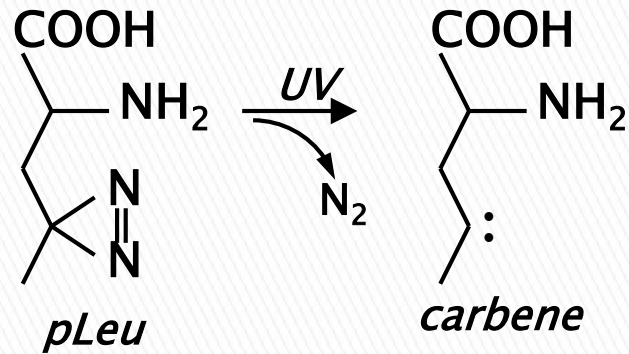
- ▶ Developed as an alternative and complement to HDX.
- ▶ Eliminates back exchange.
- ▶ Used to study protein–protein interactions (PPI) in cells.
- ▶ A variety of chemical crosslinking strategies, with triggers initiating reaction.
- ▶ Here, we evaluate photo–activatable crosslinkers in amorphous lyophilized solids.

# Photolytic labeling

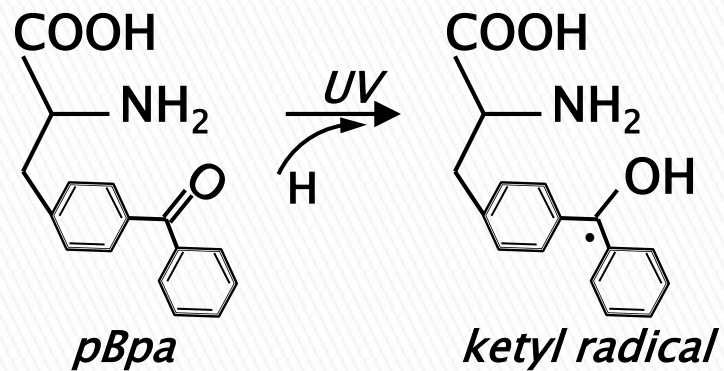


# Photoamino acids

## Photoleucine (*pLeu*)

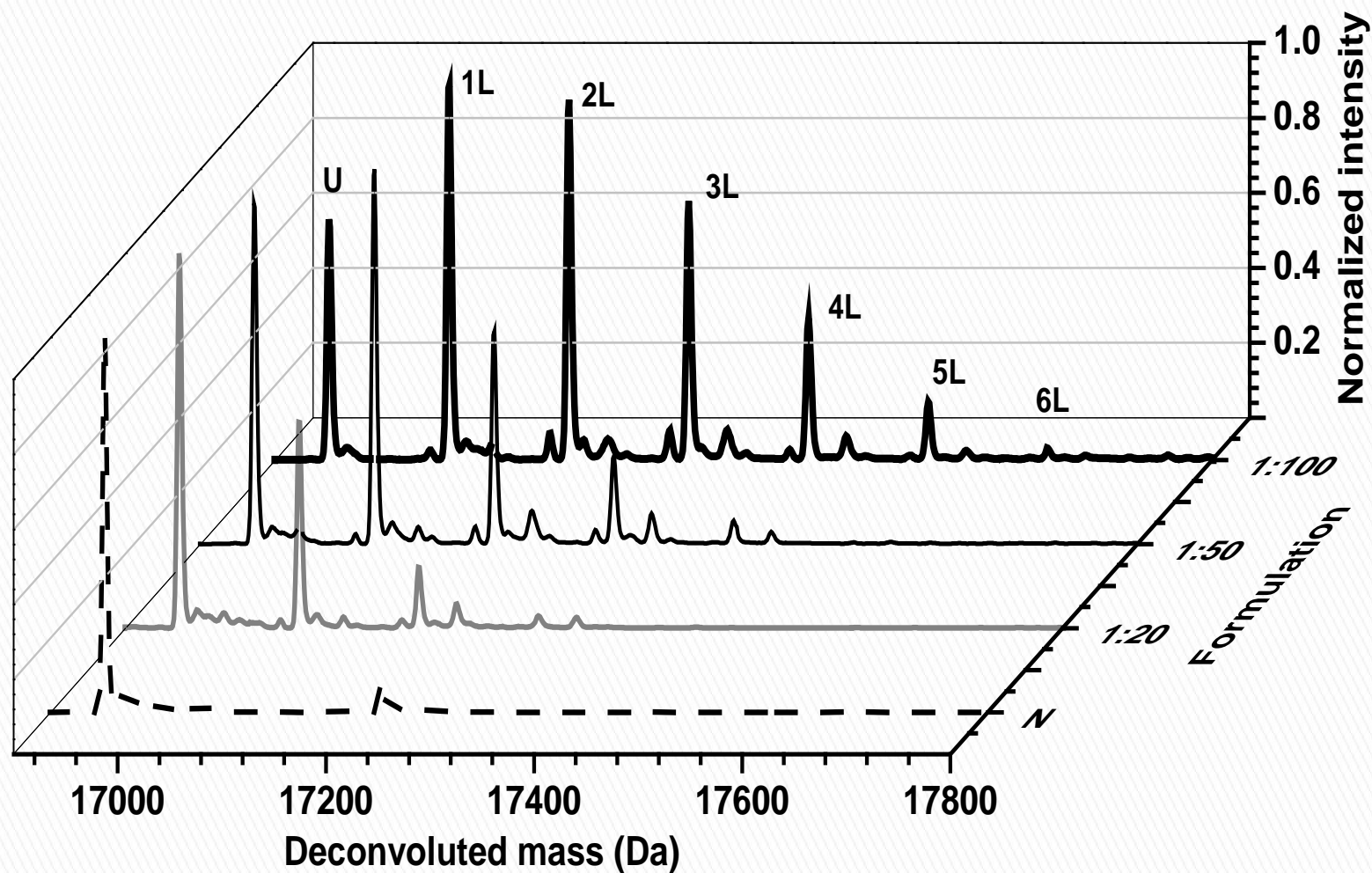


## p-benzoyl phenylalanine (*pBpa*)



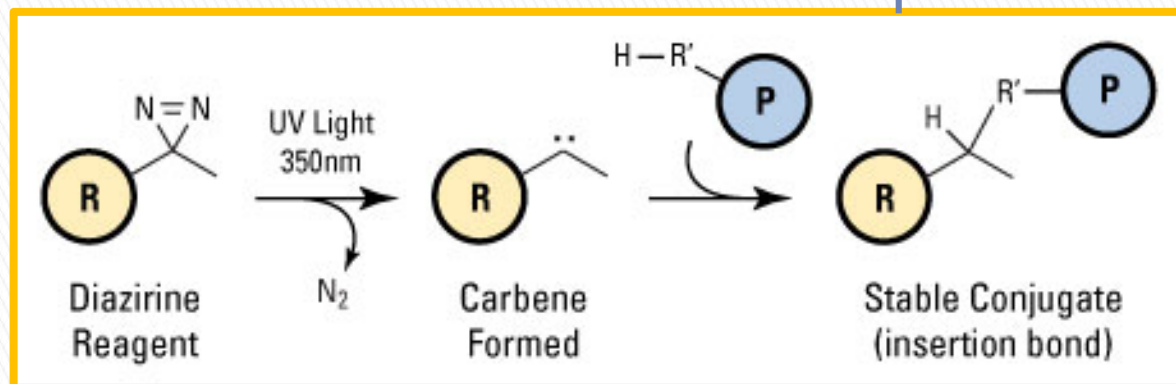
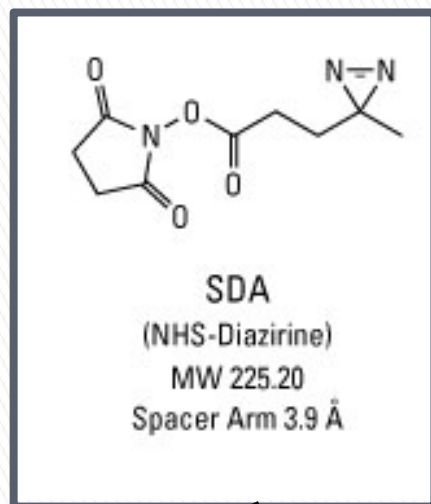
# Diazirine reacts in solids

Mb + pLeu + *sucrose* (1:2 w/w), 365 nm





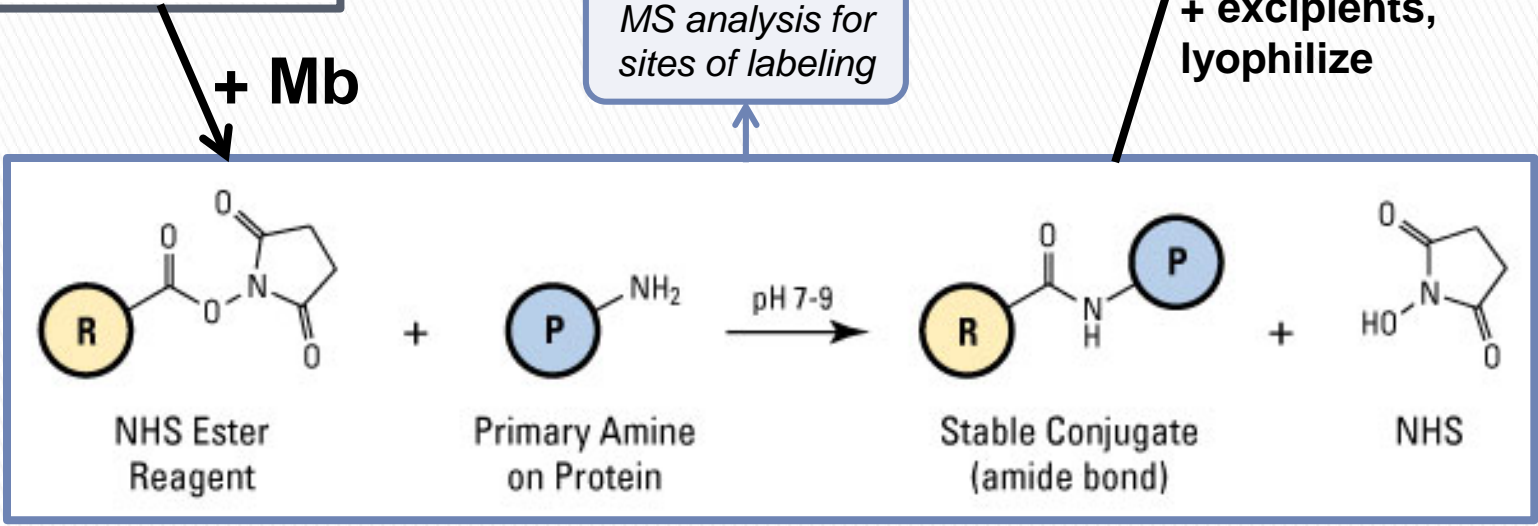
# Workflow



MS analysis for adducts formed

MS analysis for sites of labeling

+ excipients, lyophilize



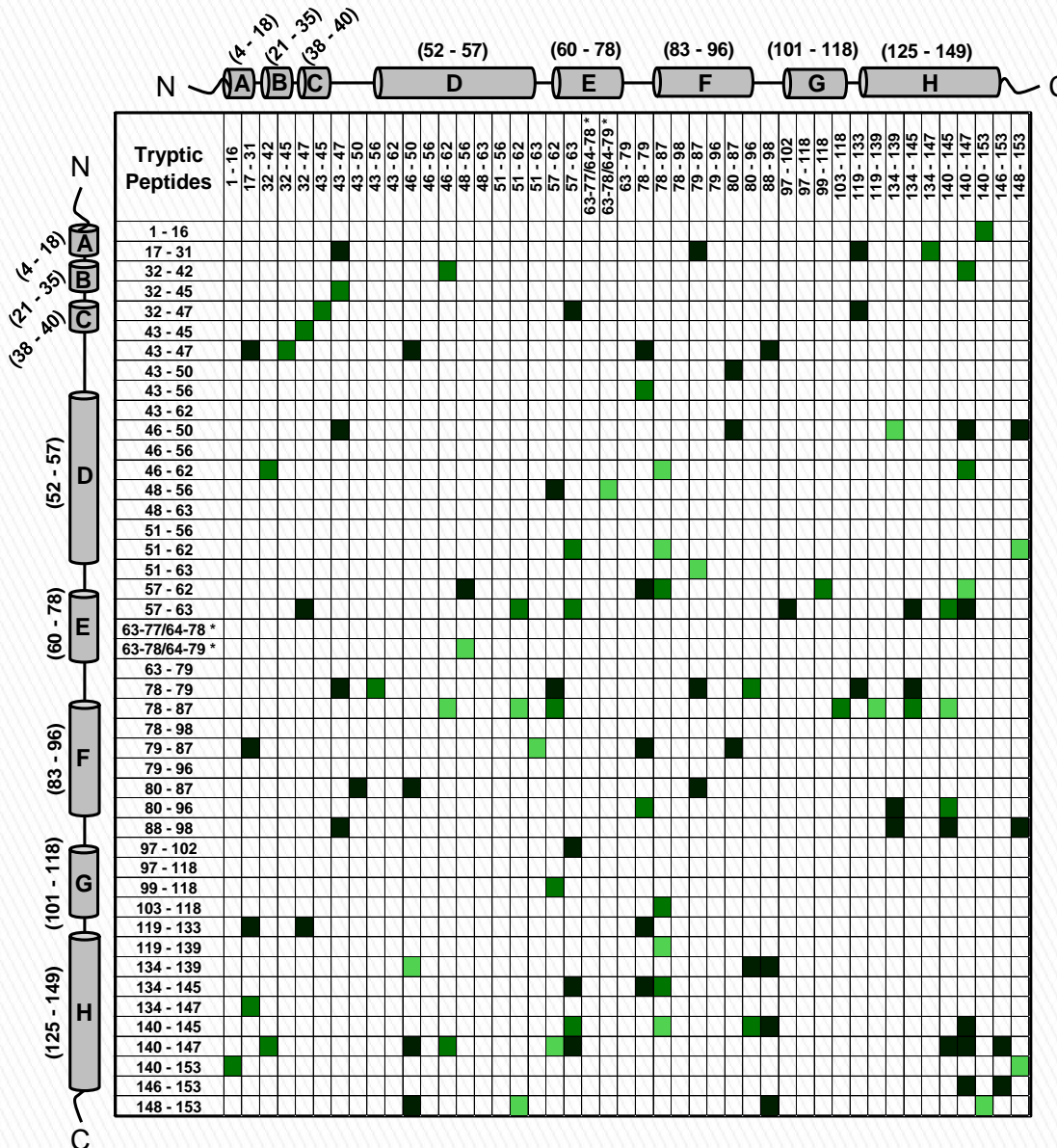
# MS analysis

- ▶ Types of adducts
  - Protein–protein adducts
  - Protein–**water** adducts
  - Protein–**excipient** adducts
- ▶ ESI LC–MS and MS/MS, trypsin digestion
- ▶ “Detection” (mass list)
  - Up to four missed cleavages (trypsin)
  - Dead–end modifications (N<sub>2</sub> loss without adduct)
  - Multiple SDA (0–4 labels per peptide)
  - Adducts of two peptides



# Protein-protein interactions

## Lyo Mb, + raffinose

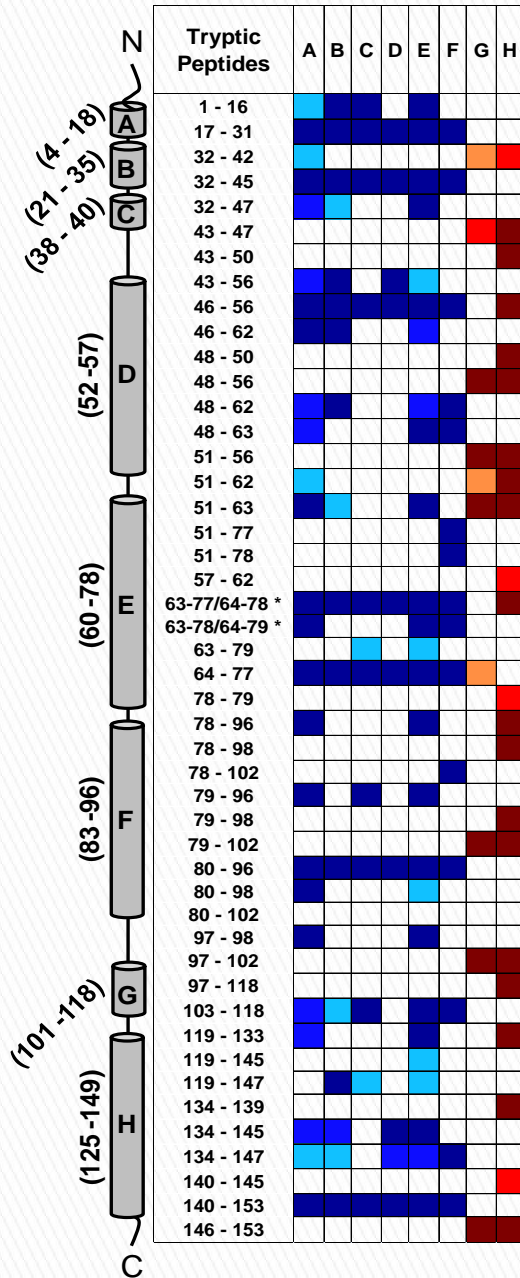


- Interactions somewhat more distributed than in excipient-free formulation.





# Water and excipient interactions



## KEY

### *Peptide/water adducts*

A – Mb alone, lyo

B – Mb alone, soln

C – Mb + raffinose, lyo

D – Mb + raffinose, soln

E – Mb + Gdn HCl, lyo

F – Mb + Gdn HCl, soln

### *Peptide/raffinose adducts*

G – Mb + raffinose, lyo

H – Mb + raffinose, soln

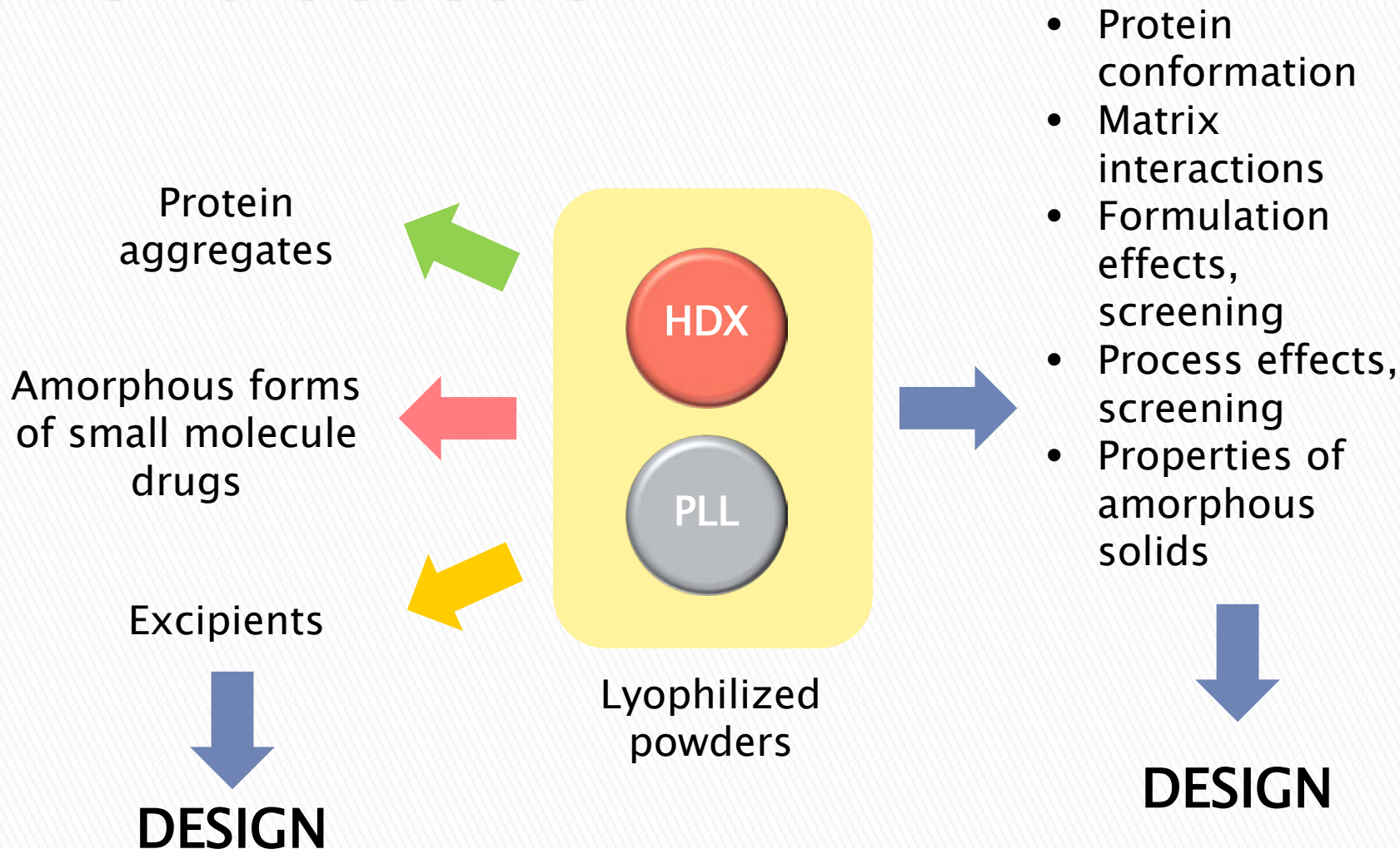


# Summary

- ▶ Photolytic labeling can be performed in solid powders using diazirine chemistry.
- ▶ SDA-labeled Mb forms adducts with protein, water and excipient (raffinose) in lyo solids.
- ▶ Formulation affects the adducts formed.
- ▶ Method allows water replacement hypothesis to be tested.



# For discussion



*“Can we have all the answers yesterday?” Hong-Ren Wang, Vertex*



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*Topp Lab, May 23, 2015*



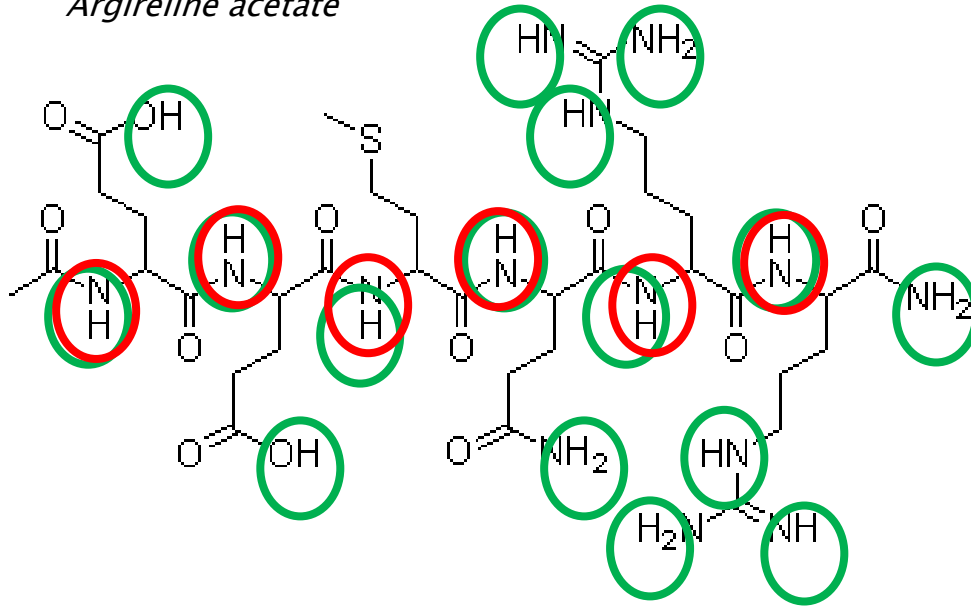
# Supplemental Slides



# Back exchange

## and the information content of HDX in solution

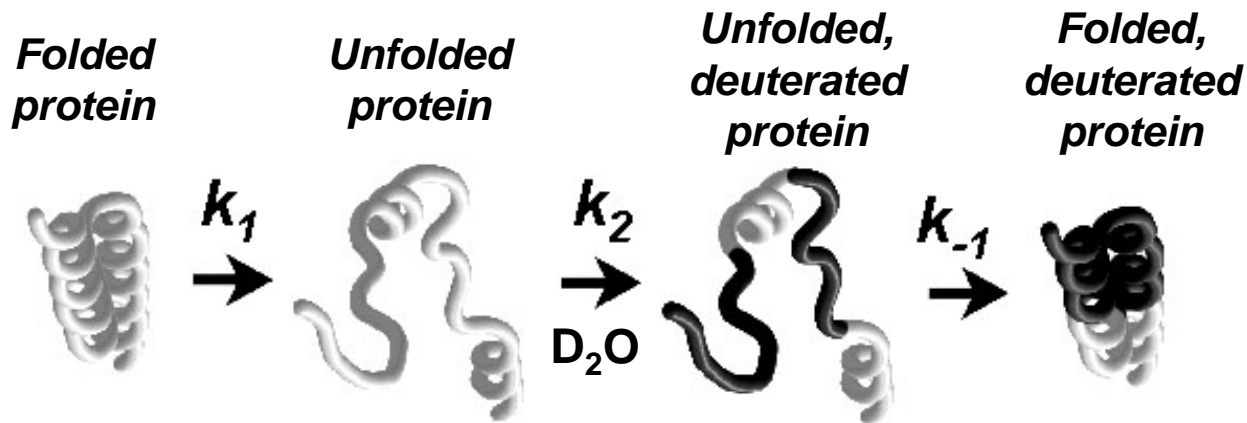
*Argireline acetate*



- ▶ HDX can occur at any labile hydrogen.
- ▶ Back exchange is rapid for side chains; only HDX of peptide bonds is detected.
- ▶ In larger proteins, HDX occurs slowly for H-atoms involved in secondary structure or buried in the core.
- ▶ HDX reports the “exposure” of the backbone to exchange.

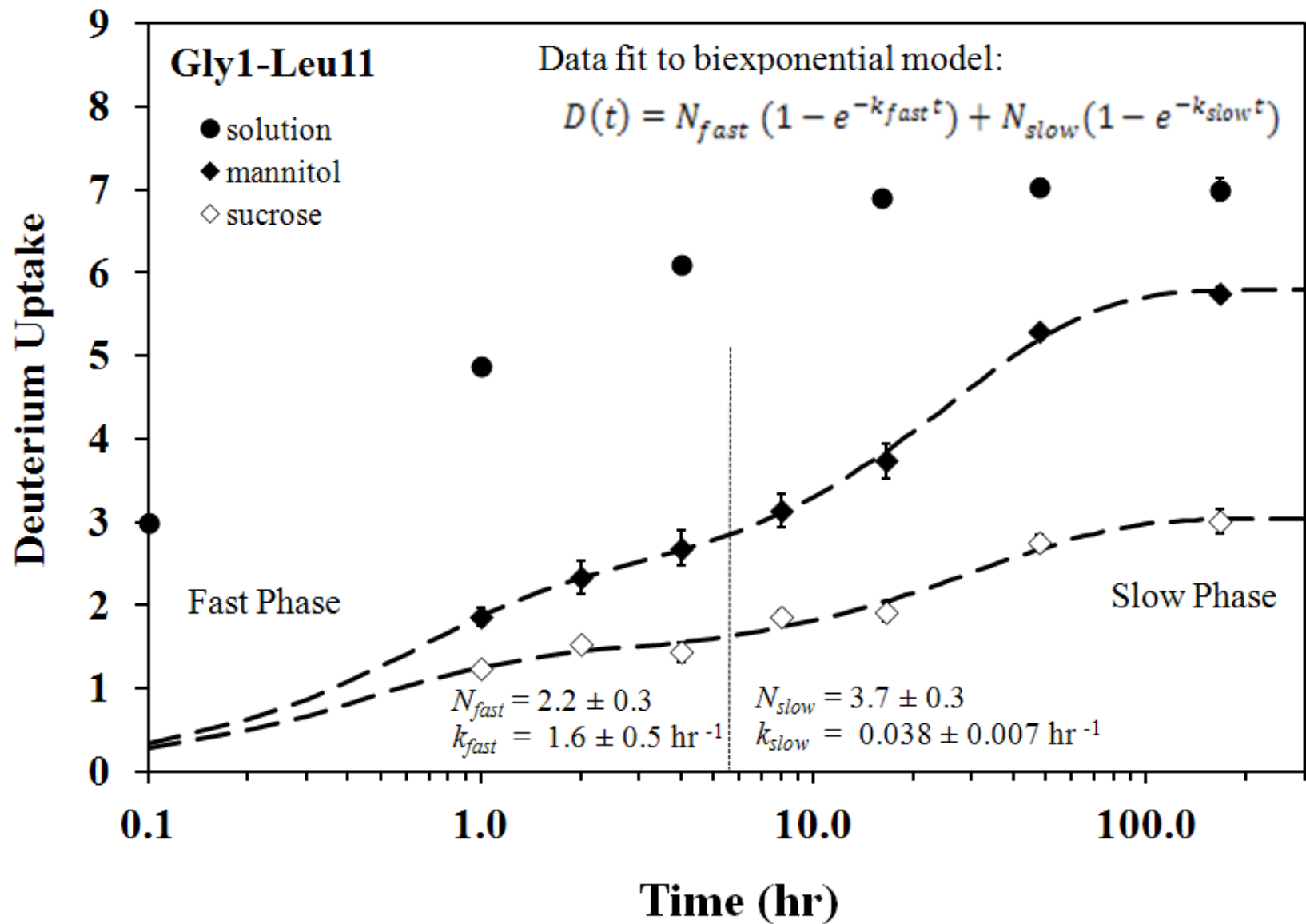
# HDX kinetics

## Protein folding and deuterium exchange



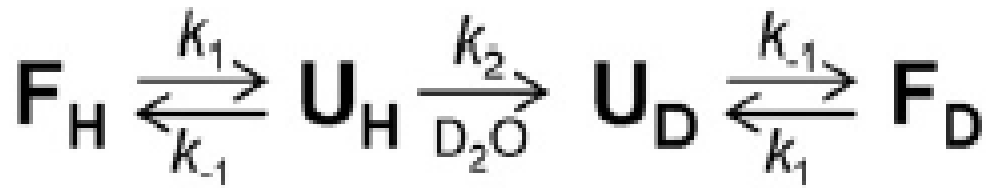
- ▶ Two kinetic processes: (i) protein unfolding and refolding, (ii) deuterium exchange.
- ▶ Two kinetic extremes:
  - Exchange much faster than refolding (EX1:  $k_2 \gg k_{-1}$ )
  - Refolding much faster than exchange (EX2:  $k_{-1} \gg k_2$ )

# Myoglobin HDX kinetics

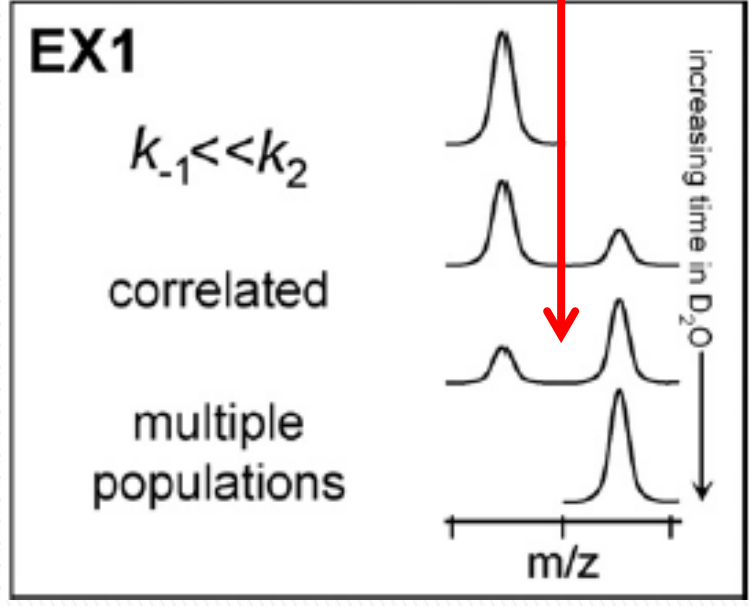
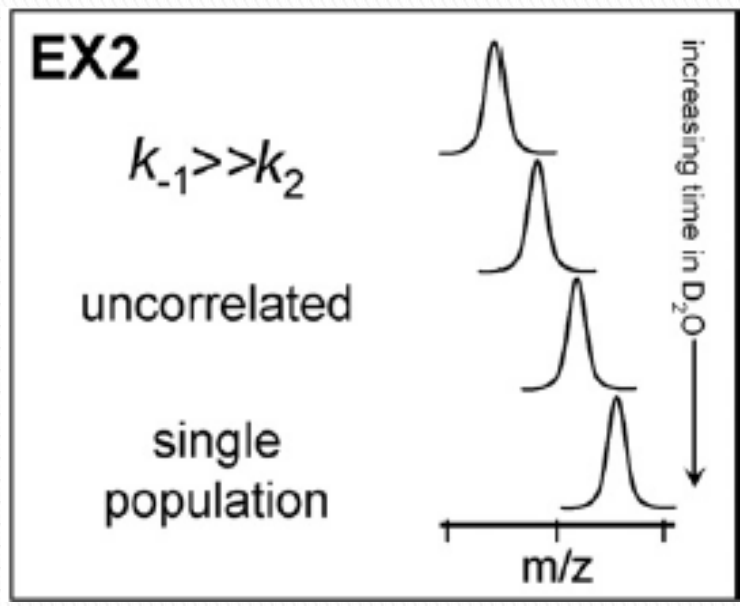




# HDX theory – EX2 and EX1 limits



Broadening of peak width may be observed instead if populations not well resolved.



most situations

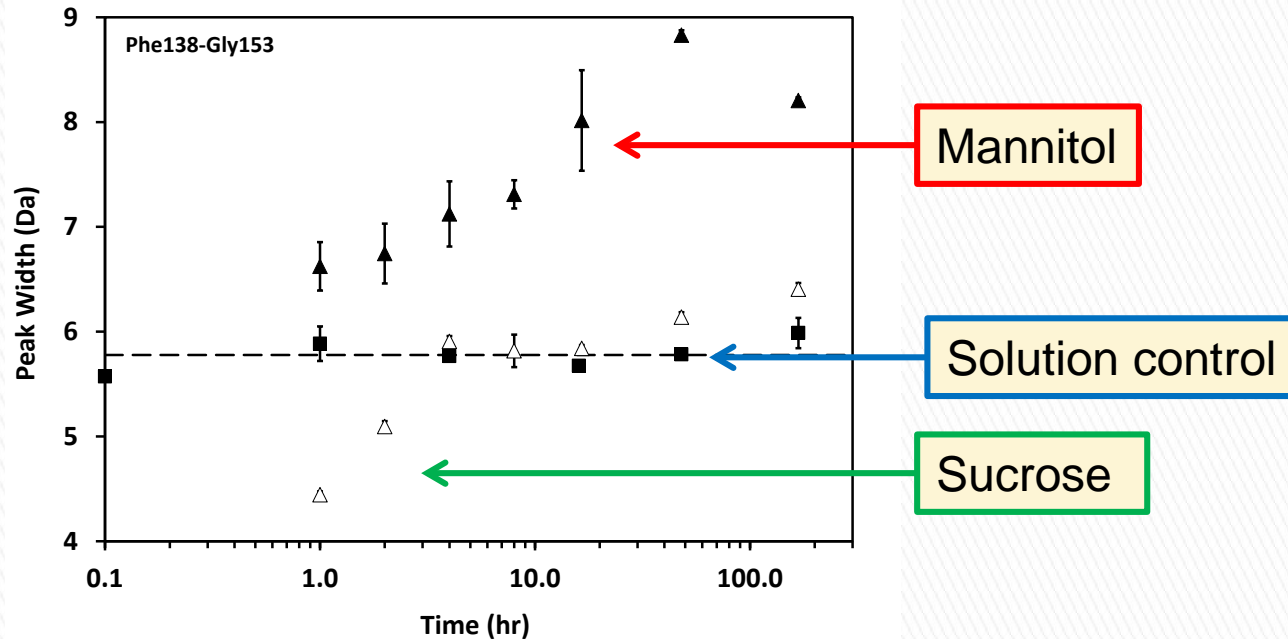
$$k_{obs} = k_1 * k_2 / k_{-1} = K_{op} * k_2$$

less common

$$k_{obs} = k_1$$

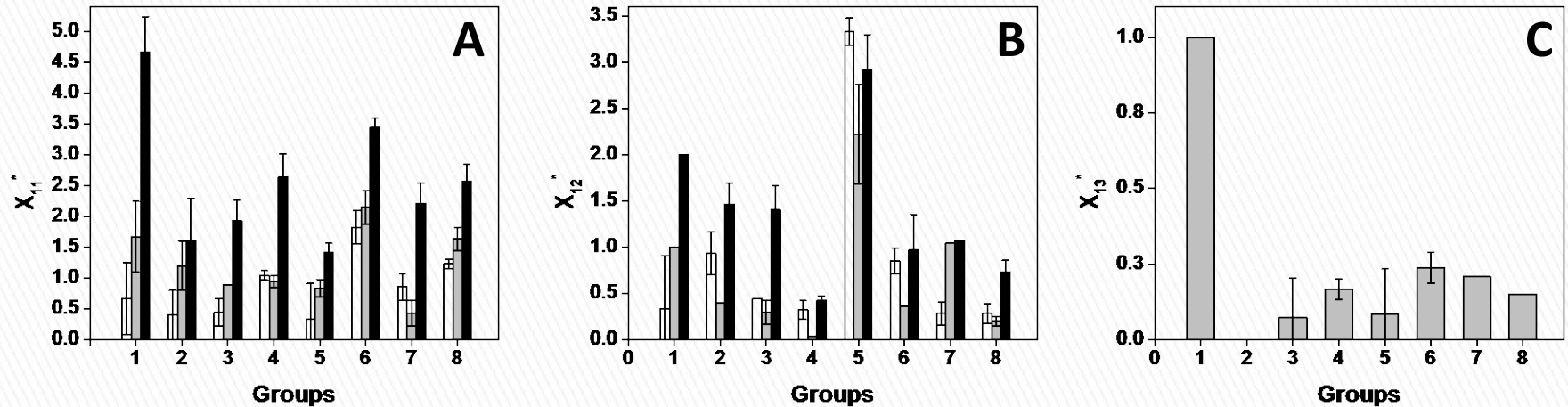
# EX2 / EX1 –like peak width broadening

Example: Phe138–Gly153 in Mb



- ▶ Greater peak widths in mannitol formulations suggest mixed EX1 /EX2 behavior, structural perturbation.
- ▶ Consistent with greater deuterium uptake overall.

# Data analysis – ssPLL



Bar graph of (A) peptide-peptide adducts, (B) peptide-water adducts and (C) peptide-raffinose adducts detected by LC-MS. White bars represent Mb-SDA lyophilized in the absence of excipients (blank), grey bars represent Mb-SDA lyophilized with raffinose and black bars represent Mb-SDA lyophilized with Gdn HCl.  $X_{1n}$  values were counted for peptides assigned to 8 groups. Bars represent mean normalized  $X_{1n}$  values ( $X_{1n}^*$ )  $\pm$  SD (n=3). Note that in the abscissa for panel (C), Group (6) spanning residues Lys<sup>78</sup>-Lys<sup>98</sup> was expanded to Lys<sup>78</sup>-Lys<sup>102</sup> to accommodate peptide Lys<sup>79</sup>-Lys<sup>102</sup> that was found to form raffinose adducts.